



# Plant nutrient dynamics: a growing appreciation for the roles of micronutrients

Sayanti Mandal<sup>1</sup> · Santosh Kumar Gupta<sup>2</sup> · Mimosa Ghorai<sup>3,4</sup> · Manoj Tukaram Patil<sup>5</sup> · Protha Biswas<sup>3</sup> · Manoj Kumar<sup>6</sup> · Radha<sup>7</sup> · Abilash Valsala Gopalakrishnan<sup>8</sup> · Vikas Moreshwar Mohture<sup>9</sup> · Md. Habibur Rahman<sup>10</sup> · Dorairaj Arvind Prasanth<sup>4</sup> · Abhijit Bhagwan Mane<sup>11,12</sup> · Niraj Kumar Jha<sup>13,14,15,16</sup> · Saurabh Kumar Jha<sup>13,14,15</sup> · Milan Kumar Lal<sup>17</sup> · Rahul Kumar Tiwari<sup>17</sup> · Abhijit Dey<sup>3</sup>

Received: 5 May 2022 / Accepted: 19 March 2023  
© The Author(s), under exclusive licence to Springer Nature B.V. 2023

## Abstract

Micronutrients are essential for plant growth and play a key role in crop nutrition. Micronutrients such as zinc (Zn), iron (Fe), copper (Cu), and manganese (Mn) are necessary for plants and humans who rely on them directly or indirectly. These minerals are important for several important cellular functions, including respiration (Fe and Cu), photosynthesis (Fe, Cu, and Mn), and transcription (Zn). Micronutrient deficiencies have gained importance as a source of ‘hidden hunger,’ focusing on iron (Fe) and zinc (Zn). Enhancing the nutritional value of staple crops seems to be a simple and effective solution. We address the methods of absorption of numerous beneficial Micronutrients, their positive characteristics, and their involvement in improving crop yield. The importance of biofortification as a procedure to enhance crop yield and as an agricultural solution to solve nutritional security challenges is discussed in this review. Biofortification boosted crop yield to relieve hidden hunger and quality parameters, proving a sustainable and cost-effective strategy. Several novel and targeted biofortification strategies for nutrient enrichment of field crops, including cereals, pulses, oilseeds, and fodder crops, have been reviewed. With the information presented here, researchers can see that biofortification promises to increase agricultural production and provide the crops with additional nutrients to ensure human food security and nutrient quality.

**Keywords** Micronutrients · Nutrient homeostasis · Nutrient acquisition · Plant nutrition · Agronomic biofortification · Biotechnological approach

## Introduction

The growth and development of agricultural plants are heavily influenced by nutrient availability; as a result, understanding the dynamics of nutrient absorption, transport, and assimilation, as well as their biological interactions, becomes more important to increasing crop productivity (Wawrzyńska and Sirko 2014). During the previous two decades, a plethora of data on plant morphological and physiological adaptations in response to variations in mineral nutrient availability has been accumulated (Krouk et al. 2011; Gruber et al. 2013; Zhao and Wu 2017; Krouk and Kiba 2020). To a certain degree, protein-coding genes involved in the absorption, mobilization, storage, and

assimilation of macro/micro-nutrients have been identified; however, regulatory networks regulating their expression in response to changes in nutritional status are still being uncovered (Liu et al. 2009; Pilon-Smits et al. 2009; Hindt and Guerinot 2012; Briat et al. 2015; Chaiwong et al. 2020). Several metabolic systems in crop plants are affected when a nutrient imbalance occurs. On the other hand, plants have developed mechanisms to deal with nutrient deficits (Hindt and Guerinot 2012; Pilon-Smits et al. 2009). Even though the soil contains many components, only 17 are now recognised as critical for crops’ normal growth and development (Bünemann et al. 2018).

Nutrients for plant growth and development are mostly derived from soils (Cogger and Brown 2016). Plants absorb the required nutrients from the soil in varying quantities depending on their requirements (Marschner 2012). An important role is played by the phyto-microbiome (plant-associated microbial population) in increasing nutrient

---

Communicated by Marian Brestic.

Extended author information available on the last page of the article

availability and crop productivity (Bhatt et al. 2020; Fendrihan and Pop 2021). In addition, agriculture and agricultural products directly impact the health of both animals and people (Brevik et al. 2020). Macronutrients are required in greater amounts by plants, while micronutrients (also known as trace elements) are required in lesser quantities; nonetheless, both nutrients play equally significant roles in plant growth and metabolism. Soil health, crop yield, the nutritional value of agricultural products, and ultimately human and animal health are all harmed when phyto-available nutrients are deficient (Shukla et al. 2018). Apart from nitrogen (N), phosphorus (P), and potassium (K) inadequacy, deficiencies of sulfur (S) and micronutrients (zinc (Zn), boron (B), iron (Fe), copper (Cu), and manganese (Mn)) have been recorded in many soils across the globe, including India (Kihara et al. 2020; Shukla et al. 2019; Piotrowska-Dlugosz et al. 2017). The essential micronutrients Zn, B, Fe, Cu, Mn, and molybdenum (Mo) are required for field crops.

Historically, various field crops that serve as a staple food on a global scale have been a key focus for biofortification. In 2004, the first biofortified crop was released in Uganda and Mozambique. The crop was vitamin A orange sweet potato variety. More than 400 varieties of 11 biofortified crops have been released for consumption in 30 countries. By 2021, almost 12 million farming households were growing biofortified crops, benefiting over 60 million people (<https://biocropshplustst.ciat.cgiar.org/about/howToDevelop>). Rice, wheat, maize, pulses, oilseed crops, and fodder crops tend to collect exogenously provided nutrients and have therefore been biofortified. So far, research on biofortified crops containing critical micronutrients such as Zn, Fe, I, Cu, and others has been extensive (Klikocka and Marks 2018). For instance, the Zn-efficient PBW 1Zn variety was introduced by PAU in 2017 and had high quantities of Zn ( $40.6 \text{ mg L}^{-1}$ ) and Fe ( $40.0 \text{ mg L}^{-1}$ ). Moreover, many biofortified crops have been released, including corn, orange, cassavas, sorghums, sweet potatoes, squash, Fe-enriched beans, Zn-enriched rice, and lentils supplemented with micronutrients. (Steur et al. 2017; Connorton and Balk 2019; Ligowe et al. 2021; Saeid et al. 2019). Deficits in micronutrients are substantial obstacles to raising the socioeconomic level of food nutrition in the twenty-first century. A straightforward method to raise the nutritional content of the average person's diet is bio-fortification (Jaiswal et al. 2022).

Genetic engineering, traditional breeding, and agronomic biofortification are current methods used to attain nutritional security (Bouis and Welch 2010; Bouis 2003). The conventional strategy entails choosing existing varieties of high-yielding crops and cross-breeding them with better nutritional content to generate staple crops with desired nutrient and agronomic features. In addition, specialized

genetic modification may enhance the concentration of micronutrients in edible plant portions (Haun et al. 2014; Michno et al. 2015; Tang et al. 2016; Dodo et al. 2008; Bonfim et al. 2007). In the present global food landscape, there is an inadequate supply of various micronutrients in dietary crops; consequently, fortifying yields with a single nutrient is insufficient to provide nutritional security, particularly if the diet is based on a narrow variety of foods. Advanced research has been conducted to determine the efficacy of applying combined nutrients to crops. Selenium and iodine biofortification of food plants was recently described by Górnaiak et al. (Górnaiak et al. 2018).

This review aims to create awareness about micronutrient delivery to plants to promote agricultural sustainability and increase yield and rectify micronutrient deficiencies in the human population. Micronutrient biofortification of staple crops for nutritional security is summarised in this review.

## Agricultural crops are a rich source of micronutrients

The dietary pattern must provide the bare minimum of important elements such as carbohydrates, proteins, lipids, vitamins, and minerals (Lean 2019). Globally, various crops are cultivated to supply this demand, including cereals, pulses, oilseeds, and fodder crops. Because developing nations lack diversified food, staple crops must include essential nutrients in an appropriate diet. Although there are more than 50,000 edible plants on the planet, just 15 of them produce 90% of the food. Two-thirds of the world's consumption of staple foods come from these three commodities: rice, maize (corn), and wheat (Shelef et al. 2017). Other important dietary intakes include millet and sorghum, potatoes, cassava, yams, dairy products, and animal items such as meat and fish. In low-income developing countries, the primary diet consists of wheat, rice, millet, corn, and sorghum (Shelef et al. 2017).

## Cereals crops

Aside from reducing global food insecurity, cereal grains can absorb more mineral nutrients in their edible part. Rice (*Oryza sativa*) is the most widely consumed cereal grain by more than half of the world's population and accounts for more than 42% of calorie consumption (Huang et al. 2020). Rice is the fastest-growing staple food in Africa and Latin America, accounting for about 90% worldwide consumption. Rice contains various vitamins (such as vitamin B1, B2, B3, B6), phyto-molecules, and minerals such as sodium, zinc, iron, copper, potassium, magnesium, and phosphorus.

Wheat is the second most common food after rice in South Asia, Turkey, and China, accounting for more than 70% of daily calorie consumption. As a result, its nutritional content is critical for ensuring nutritional security. The testa, aleuronic layer, embryo, and endosperm are all parts of wheat grain. The embryo is the live component of the grain, and it has a high concentration of nutritional value. While the endosperm is largely starch, the aleuronic layer contains most nutrients (such as protein, minerals, and vitamins) (Rosa-Sibakov et al. 2015). Its versatility adds to its value, as its seeds may be processed into semolina, flour, and other fundamental components for pasta, bread, and pastry items. In addition to wheat and rice, corn, also known as maize, is the world's most important staple grain. Most of its popularity comes from the fact that it is a good source of food for both human and animals. It may be prepared in various methods, including boiling, roasting, frying, grinding, and fermenting, and is often used in gruel, bread, cakes, porridges, and alcoholic drinks. In industries, its use is in sweeteners, oils, food thickeners, and non-consumables. Its nutritional composition comprises 72% carbohydrate, 10% protein, and 4% fat. Except for vitamin B-12, maize includes a variety of essential vitamins (Rosa-Sibakov et al. 2015). Maize's germ and endosperm contain 80% and less than 1% of the total mineral content. In maize, the most common minerals are K, Mg, Zn, and P (in the form of phytate). Methionine and cystine are two sulphur-containing amino acids. In the state of methionine and cysteine, sulfur may be found in the body. Ca and Fe are present in negligible amounts, and other trace minerals include Mn, Se, Cu, and I (Nuss and Tanumihardjo 2010).

## Pulses crops

Pulses are also known as a “superfood” because they have a lot of nutrients that can help you stay healthy. The FAO has designated nutrition security as its 2016 topic, and this shows that they acknowledge the importance of achieving this goal (Food and Agricultural Organization of the United Nations). After cereals, they are the second most important crop in the agricultural sector, with a large consumption (over 70%) of lentils, peas, beans, and chickpeas globally (Callens et al. 2019). They are also important for their social and economic impact because of their inter-cropping systems. Cereals have a significant socioeconomic influence because of the intercropping systems, including low fertilizer and water needs, great disease resistance, extended storage duration, and the capacity to adapt to extreme environments (Brueck and Lammel 2016). These characteristics, when combined, possess technological/functional features with broad industrial relevance for the production of functional foods. In

recent years, six pulses, namely chickpea (*Cicer arietinum*), field pea (*Pisum sativum*), lupin (*Lupinus*), faba/broad bean (*Vicia fabae*), lentil (*Lens culinaris*), and mung bean (*Vigna radiata*), have acquired prominence owing to the large amounts of these crops produced worldwide (Callens et al. 2019). India is the largest producer in Canada, Myanmar, and China.

Furthermore, they are high in protein (up to 30% of their dry weight in protein), carbohydrates, dietary fiber, and calories. They are also high in important bioactive compounds and necessary minerals and vitamins needed by the human body (Rebello et al. 2014). Lentils and beans are the pulses with the highest Fe and Zn content, followed by peas and lentils. Beans often have the greatest concentrations of iron and other elements. It is, nevertheless, vital to remove phytate and other inhibitors since they reduce the digestion and bioavailability of minerals. Pulses, owing to their high mineral concentration, would become potential sources of Zn and Fe after decomposition (Venkidasamy et al. 2019). Although cooked seeds maintained about 70–75% of water-soluble vitamins, raw bean samples contained 0.49 mg vitamin B12 and 0.30 mg folic acid. The folate concentration of beans accounts for 95% of daily requirements.

## Oilseed crops

Oilseed crops are generally planted to extract the oil contained inside the seeds. Vegetable oils are widely utilized in domestic cooking and other food products such as baked goods and snacks worldwide. Additionally, they serve as raw materials for a variety of oleo-chemical businesses. Canola, safflower, maize, sunflower, olive, soybean, and peanut crops are examples of oilseed crops. Oilseeds are a key vegetable oil source with greater production of oil when compared to other crops (Zafar et al. 2019). These crops are especially essential among low-income households in semi-arid tropical areas since they account for 40% of their total calorie consumption. Recent studies have demonstrated that rape oilseeds contain significant protein and minerals such as Zn, Fe, Ca, and other trace elements, making them a viable alternative to conventional crops for addressing nutritional stress (Kowalska et al. 2020). Groundnut, soya bean, castor bean, and other temporary oilseed crops are separated from permanent oilseed crops such as coconut, oil palm, and olives. Permanent oilseed crops include coconut, oil palm, and olives. Soybean, which is readily accessible as a meal and is high in amino acids, is a commodity that is extensively traded across the world. Because of their low lignin level and high mineral and nutrient content, soybean hulls are an excellent fibre, protein, and energy source. Proteins, amino acids, vitamins, and minerals are included in canola

meals with a high sulphur concentration. Hemp seed and mustard seed oil are fortified with good nutritional characteristics and minerals such as Ca, Mg, P, S, K, Fe, and Zn.

Similarly, various oilseeds are very nutritious for humans. Because of their high oil content, brassica crops, including mustard, canola, and rape, are receiving much attention in the bio-diesel business for creating feedstock oils. The large levels of phytic acid and other binding agents in oilseeds, on the other hand, restrict mineral bioavailability from the seeds. As a result, phytate removal methods are being developed to manage or eliminate mineral binding in oilseed products. Oilseed supplemented with minerals by biofortification, on the other hand, may help to reduce the mineral shortage. For example, under acidic conditions, foliar application of  $\text{Na}_2\text{SeO}_4$  enhanced the Se concentration in the plant components of oilseed rape (Szákóvá et al. 2017). However, the biofortification of oilseed crops has received little attention, and this area needs to be investigated more to improve their nutritional value.

## Mineral acquisition and transport via molecular mechanisms

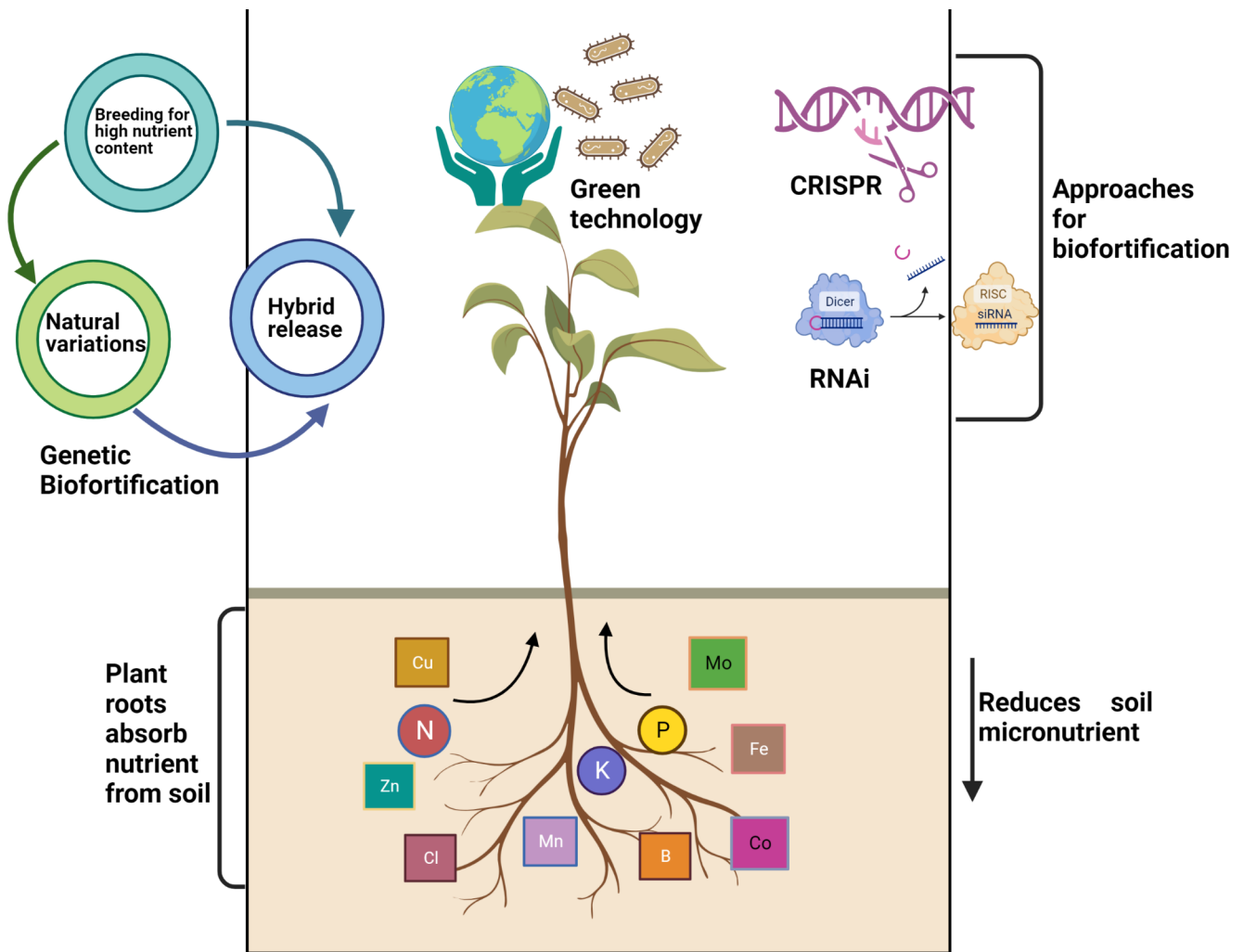
Plants require micronutrients. The absorption of micronutrients from the soil, diffusion throughout the plant, delivery to sink organs, and allocation to particular metalloproteins in plants are all optimized via a highly specialized and tightly controlled network in the plant. *Arabidopsis thaliana* and rice have been the primary models for studying the molecular processes of plant micronutrient absorption and usage efficiency (Sperotto et al. 2014; Xue et al. 2016). In the future, the knowledge gleaned from these models will need to be applied to other agronomically important crops.

Mineral uptake, transport, and accumulation in crop plants must be well understood to enhance nutritional bioavailability. These processes are probably regulated by genes, many of which are unknown. Several studies have found genes involved in translocation into numerous vegetative tissues and eventually to seeds (Sperotto et al. 2014; Jeong et al. 2017). But the genes expressed by phloem involved in mineral loading and mobilization to various sink tissues are only known to a very limited extent (Braun et al. 2014). As a result, although research on particular transporters may help us understand their function, whole-plant studies are needed to identify the most important transporters for seed mineral delivery. Mineral acquisition and mobilization in plants have been extensively researched (González-Guerrero et al. 2016; Xue et al. 2016). However, numerous stressors may result in the non-availability of critical nutrition elements, impairing crop development.

## Uptake of micronutrients

The molecular processes of plant absorption of most micronutrients have been explored in recent decades (Connorton et al. 2017; Stanton et al. 2021). The involvement of rhizosphere acidity in acquiring metal micronutrients is well established. It has recently been shown that root exudates play an important role in mobilizing dicot micronutrients (Fig. 1.). While it was previously known that cereals acquire Fe through the secretion of phytosiderophores into the rhizosphere, the central role of secreted coumarins in mobilizing Fe in the vicinity of growing roots has been highlighted in several dicot species, with possible implications for the acquisition of other micronutrients and the shaping of the bacterial communities associated with the roots (Tsai and Schmidt 2017; Stringlis et al. 2018). These are known as strategies I (proton extrusion, reduction, and absorption of Fe (II)) and II (phytosiderophores). Additionally, the YSI (Yellow Stripe Oligopeptide Transporters 1) transporter transports the ferric-phytosiderophores complexes from the rhizospheres into plant roots (Saini et al. 2016). Furthermore, current research has shown the presence of unique processes for metal absorption from arbuscular mycorrhizal fungi, which are particularly important in natural ecosystems and organic agrosystems (Senovilla et al. 2020). Up to 50% of plant Zn absorption is attributed to root mycorrhizal colonisation (Watts-Williams et al. 2015). Reducing root mycorrhizal colonisation resulted in significant cadmium (Cd) build-up in wheat (Yazici et al. 2021). Thus, it is critical to pay close attention to the number and activity of mycorrhizal fungi in agricultural soils to control micronutrient nutrition and prevent Cd build-up in crop plants (Ma et al. 2021).

Even though plants need micronutrients, too much of them is often toxic and causes harmful things to happen (Raffa et al. 2021), such as a decrease in photosynthetic pigments (Aslam et al. 2021), a decrease in the integrity and permeability of membranes (Stoyanova and Doncheva 2002), an increase in oxidative stress caused by the production and accumulation of reactive oxygen species (ROS), as well as an increase in the activity of antioxidant enzymes (Balafrej et al. 2020), and in the worst cases, cell death (Zhang et al. 2022). Excess micronutrients have negative consequences such as diminished photosynthetic pigments, oxidative stress, changes in membrane integrity and permeability, suppression of protein function, interference with absorption and use of other mineral elements, stunted development, and lower yields (Zhang et al. 2022; Raffa et al. 2021).



**Fig. 1** Schematic illustration of micronutrients absorption in plants and different methods of biofortification. The absorption of micronutrients by plants results in the depletion of the soil micronutrients. Plants are

deficient in micronutrients that can be biofortified through various approaches. Figure created with BioRender (biorender.com)

## Distribution of micronutrients within the plant

Micronutrients are delivered to sink organs inside the plant through the sap, often the youngest leaves and seeds. This is accomplished by the movement of nutrients across membranes and the presence of tiny organic ligands that preserve the nutrients' solubility in solution and avoid mis-metallation (Olsen and Palmgren 2014; Connorton et al. 2017). While numerous genes involved in micronutrient mobilization and absorption from the soil have been discovered, the processes for loading micronutrients into the xylem and phloem and the ligands in conducting tissues are still largely unknown micronutrients. The transporters in the nodes and tiller buds of plants play important roles in the distribution of micronutrients in plants, especially in the delivery of micronutrients to the younger leaves and seeds, and should

be studied further (Shao et al. 2018; Mu et al. 2021). Only a rudimentary understanding of the mechanics of micronutrient distribution to the edible organs exists (Andresen et al. 2018). To breed or engineer plants for targeted micronutrient allocation, a deeper understanding of the processes of micronutrient distribution to the organs and tissues would be required.

Additionally, metal recycling and reuse are mainly ignored during the shift from vegetative to reproductive development. According to preliminary findings, metal remobilization from senescing leaves looks to be a viable area for modulating metal accumulation in seeds (Mari et al. 2020). Wheat or Arabidopsis leaves with autophagy-mediated modification of leaf senescence and nutrient recycling change the levels of protein and mineral micronutrients in the seeds (Chen et al. 2019; Pottier et al. 2019). In addition, the method of mineral micronutrient loading into seeds is



another important step that has to be investigated further. There is only a limited amount of information available on the penetration of micronutrients into seeds. Metal-pumping ATPases have been shown to play a crucial role in releasing Zn from the mother tissues before its entrance into the seed (Olsen et al. 2016). The production of ascorbate is utilized to lower Fe levels in the bloodstream and enable it to be absorbed by the embryo (Grillet et al. 2014). This study will need extensive characterization of the dynamics of micronutrient distribution in plant organs and tissues through the use of basic imaging methods and metal isotopes as tracers need to be developed further (Sheraz et al. 2021).

## Micronutrient interactions

Aside from their relevance for nitrogen metabolism, micronutrients interact significantly with several macronutrients. Indeed, deficiency symptoms are often caused by dietary imbalances rather than a deficit of a specific micronutrient. Homeostasis of micronutrients and other mineral nutrients, such as inorganic phosphate (Pi) with Zn and Fe, have essential interactions (Dong et al. 2017; Hanikenne et al. 2021; Watts-Williams et al. 2015). On the other hand, Zn deficiency promotes Pi absorption, whereas root Zn absorption is inhibited by high Pi levels (Kisko et al. 2018). The uptake of Zn and Fe by the roots and their transport and remobilization from the roots to the shoots are positively affected by nitrogen fertilization (Kutman et al. 2011). In *Arabidopsis*, Iron-Regulated Transporter or Zrt-/Irt-like Proteins (IRT/ZIP) family and Yellow StripeLike (YSL) were identified (Kobayashi et al. 2019). Following this, the concentrations and distributions of zinc, iron, and proteins in seeds are highly connected (Cakmak et al. 2010). On the other hand, sulphur (S) fertilization has a detrimental effect on Mo absorption, resulting in Mo deficiency in plants (Maillard et al. 2016a, b). The metalloproteins involved and their biological functions must be explored to establish the molecular underpinnings of these interactions.

Enhancement of the accumulation of chemically identical hazardous trace elements is a key danger faced in biofortification techniques targeting mineral micronutrients (Kawakami and Bhullar 2021). Enhancing the expression of a transporter involved in Cd sequestration in root vacuoles or knocking down a gene involved in Cd absorption in roots have been effective techniques for preventing Cd accumulation-up in grains in rice. (Ueno et al. 2010; Ishikawa et al. 2012). Due to their chemical similarity, micronutrient metals compete with non-essential metals for transport and binding to ligands. It is necessary to have an extensive biochemical understanding of the substrate specificities and relative affinities of proteins and transporters for micronutrients and

hazardous substances with comparable chemical characteristics to address these essential scientific problems in detail (e.g., Zn and Cd). To enhance the exclusion of non-biogenic metals, the factors of metal specificity, especially among divalent metals, must be well characterised (Pottier et al. 2015).

## Specification of micronutrients

In plants, the most important metal ligands, like citrate or nicotianamine, and their involvement in micronutrient transport across cells and organs have been well established (Clemens 2019). For example, the bioavailability of Fe and Zn in rice grains was improved by increasing the concentration of the metal-binding molecule nicotianamine by activating the nicotianamine synthase gene (Lee et al. 2011). However, micronutrient speciation and subcellular distribution processes must be better understood to improve micronutrient content in edible crop plant parts and optimize metal distribution for photosynthetic and N metabolism. Identifying the metalloproteome and metallome for each essential metal using analytic and spectroscopic approaches and examining micronutrient speciation is critical to achieving this aim (Flis et al. 2016). Because metal speciation occurs in various cell parts, understanding the processes that govern micronutrient allocation is essential for controlling metal speciation. For instance, in the vacuole, Fe is generally stored as an insoluble compound with phytate, but in the plastids, it is stored as bioavailable ferritin (Mari et al. 2020).

## Micronutrients in soil: global status

In many parts of the world, micronutrient insufficiency in soil has been tested using different analytical methods. High-yielding crops require many fertilizers, but there is not enough micronutrient supplementation to make up for it. Deficiencies of Zn and B were found to be 49% and 31%, respectively, in major regions of the world's soil (Shukla et al. 2021). Other mineral shortages were 15%, 14%, 10%, and 3% for Mo, Cu, Mn, and Fe. Boron, which is found in soils in the form of  $\text{BO}_3^{3-}$ , is important for mechanical resilience and cell membrane strengthening. Low B availability in soil indicators has been found in a few places in practically every nation, notably in Nepal, India, the Philippines, and Thailand. Cu well-known function as a catalyst is involved in the production of proteins and vitamin A and the activation of enzymes in a variety of plant-growth processes. Copper toxicity has been reported in soil samples from Brazil, the Philippines, Tonga, and Italy, among other places.

Iron (Fe) is well recognised as a component of haemoglobin and various other enzymes involved in nitrogen fixation, energy transmission, and lignin formation, among other processes. In Mexico, Malta, Turkey, and a few other nations' areas, there is a deficiency in Fe bioavailable form in soil. A different micronutrient, called Mo, is used to make nitrogenase, a key enzyme in the nitrogen fixation process in the rhizobium legume symbiosis (Lešková et al. 2017). In places with acidic soils, such as Africa (Zambia, Sierra Leone, Ghana, Nigeria), Nepal, Brazil, and New Zealand, Mo deficiency is more common. For the most part, the significance of the micronutrient Mn in photosynthesis and nitrogen metabolism is well-known. Still, it also helps plants use more P and Ca by increasing the number of water molecules that may be photolyzed (Lešková et al. 2017). Mn deficiency is found in alkaline soils, while Mn sufficiency is seen in acidic soils, indicating that soil pH influences Mn availability. A high Mn deficit has been recorded in alkaline soils in Syria, India, Egypt, Pakistan, Italy, and Lebanon. Even though Zn insufficiency has been reported regularly around the globe, it is more widespread in the following countries: Iraq; Pakistan, India; Turkey; Syria; Mexico, Italy, Lebanon, Nepal, Tanzania, and Thailand (Singh 2001).

Micronutrients are found insufficient concentrations in most Indian soils; however, their bioavailability for plant absorption is limited; as a result, soils in numerous Indian locations are deficient (Singh 2001). Indian grounds have a wide range of boron deficiencies, from 68% in red soils of Bihar to 2% in alluvial soils of Gujarat. Due to heavy rainfall, which causes a reduction in water-soluble B, maximum B insufficiency (54–86%) was recorded in the Alfisol soils of West Bengal and Assam. In the Indo-Gangetic plains with salty soil, there was a greater concentration of B, while the state of Rajasthan and Madhya Pradesh had a moderate amount. In addition, it was discovered that Kerala, the Himalayan Tarai zone, Bihar, Uttar Pradesh, and north Madhya Pradesh have copper-deficient soils (Singh 2001). Nutrient deficiency impairs the ability of the photosynthetic system to operate (Morales et al. 2020). Under a variety of stress conditions, a low stomatal aperture reduces photosynthesis. As a result, it is often observed in reaction to drought and extensively recorded in compacted soil-grown plants or soils causing deficits in the macronutrients K and P, as well as the micronutrients Zn and Cu. There is evidence of altered chloroplast ultrastructure and function in particular situations, such as plants lacking in K or micronutrients (Morales et al. 2018).

The alkali soils of the Indo-Gangetic area have low levels of plant-available iron, although iron is found in greater quantity in the earth's crust than in its soils. According to soil study studies, around 12% of Indian soils are deficient

in Fe. According to research, manganese-deficient soil is very unusual in India, with approximately 1–5% of surface soil samples being confirmed to be Mn deficient. Manganese deficiency was prevalent in Haryana, Bihar, Punjab, and Madhya Pradesh, among other places. Furthermore, excessive lime treatment in red lateritic soils of Orissa resulted in Mn shortage (Singh et al. 2006). More than 11% of the soil in India doesn't have enough molybdenum. This is true for all types of soil, from hill soils in Andhra Pradesh to the coast of India's Malabar region and the north and northeast Himalayas (Bhupalraj et al. 2002; Singh et al. 2006).

Conversely, calcareous alkaline soils in the Punjab area have high accessible Mo concentrations, and as a result, possible toxicity in crops has been documented in several crops. Wheat straw, as fodder, was discovered to have a high Mo concentration and was consequently harmful to cattle. In India, 51.2% of soil samples were Zn deficient, making Indian soils the world's most deficient throughout Zn, with a widespread deficiency in the Indo-Gangetic plains. States like Haryana, Punjab, Uttar Pradesh, Bihar, Andhra Pradesh, and Madhya Pradesh have shown a decrease in Zn deficiency (Bhupalraj et al. 2002; Singh et al. 2006).

## Micronutrient enrichment in field crops includes several approaches

Biofortification raises the nutritional quality of a target crop by increasing the concentration of micronutrients in edible sections without compromising agronomic traits such as insect resistance, yield, or drought tolerance (Klikocka and Marks 2018; Murgia et al. 2013). Conventional biofortification, transgenic, and agronomic biofortification are three key ways to achieve nutritional security via biofortification (Waters and Sankaran 2011). The traditional approach is to select existing high-yielding crops with greater nutrients of interest and cross-breeding using standard procedures to generate staple crops with desired nutritional and agronomic qualities. Specific genetic modification may increase the concentration of micronutrients in edible plant portions, resulting in genetic biofortification. To boost the micronutrient content of food plants, non-genetic approaches like agronomic biofortification may be more effective. This method boosts grain yields and increases nutritional quality (Cakmak and Kutman 2017; Khoshgoftarmanesh et al. 2010). Using N, Zn, and Fe fertilization, Kumar et al. described the agronomic intervention as a sustainable and cost-effective technique to promote plant development, yield contributing characteristics, and Zn, Fe content in rice crops (Kumar et al. 2018).

Agronomic biofortification refers to the application of micronutrient fertilizers to the soil and/or plant to enrich

the edible component of the field crop with micronutrients (Cakmak and Kutman 2017). Numerous biofortified crops, such as cereals, pulses, oilseeds, fodder crops, and others, have been grown across the globe till today. This technique gives the necessary nutrient externally before or during plant development. Foliar treatment improves the nutrient content in the leaf for transmission to other plant parts, while soil application maintains an adequate amount of food for root absorption. When applied to the soil, nutrients are ensured for root absorption, and however, when applied topically, nutrients are increased in the leaf, which may then be transported to other plant parts. The availability of the micronutrient and hence the success rate of biofortification is influenced by soil qualities such as pH, calcareous nature, etc. Therefore, the availability of micronutrients and the success rate of biofortification are influenced by soil qualities such as pH, calcareous nature, and other factors. Thus, the mineral enrichment in a given crop is affected by fertilization application technique, soil conditions, and additives.

## Traditional approach

### Agronomic biofortification

In this method, biofortification of crops (legumes and cereals) through foliar spray application of micronutrients. This application helps acquire more nutrients in reproductive parts, and hence healthier and natural biofortified grains are harvested. Micro nutrients are applied in liquid form in aerial parts of plants. After application, these micro nutrients are absorbed through stomata and epidermis, and finally become part of the food chain. Minerals such as selenium, zinc, calcium, etc., are supplied to crops alongside irrigation and are readily available for uptake. This results accumulation of these micronutrients in eatable parts of the plants. For the application of minerals through soils at the time of fertilization results perfect biofortification of crops that are usually completed through NPK. These are applied in the soil bed before sowing or seed using different seed cum fertilizer drills. In this way, they are absorbed and made part of the food chain through root uptake. Microbe-mediated enhanced uptake of nutrients for biofortification of vegetables and crops are also established techniques. Different microbial species, i.e., rhizobium bacteria, mycorrhizae fungi, etc., help plants in nutrient acquisition through mutualism.

### Green technology (bacterial and endophytic biofortification)

Microorganisms are used in green technology to enhance the soil nutrient status by increasing nutrient accessibility to the crops. According to research, excessive usage of chemical fertilizers has negative consequences for the environment and human health. As a result, microorganisms (*Trichodesmium*, *Azospirillum* spp., *Pseudomonas* spp., *Mycorhiza*, *Bacillus* spp. *Enterobacter*, *Azoarcus* spp. etc.) harbouring biofertilizers may be a cost-effective and environmentally friendly alternative to chemical fertilizers for plant development and soil fertility (Fig. 1.). Plant growth-promoting microorganisms benefit plant health in various ways, including biological nitrogen fixation, the generation of numerous plant growth hormones, hydrolytic enzymes, siderophores, HCN, and the solubilisation of K, Zn, and P (Kaur et al. 2020). Bioavailability of nutrients cannot be achieved entirely by intrinsic plant-based mechanisms, including chelator secretion, organic acid synthesis, and phyto-siderophores. Microorganisms create chelating chemicals that form complexes with Zn and release them at the root surface, increasing Zn availability and resulting in Zn biofortification in plants. There have been several research on the positive impact of microorganisms in crop biofortification. Bacteria and fungus are the most common sources of biofertilizers.

### Bacterial biofortification

The introduction of *Pseudomonas* and *Bacillus* sp. converted soluble zinc complex chemicals ( $ZnS$ ,  $ZnO$ , and  $ZnCO_3$ ) into zinc ions in broth culture (Prahara et al. 2021). The application of *Arthrobacter* sp. DS-179 and *Arthrobacter sulfonivorans* increased the qualitative and quantitative parameters linked to Zn and Fe concentrations in wheat crops (DS-68) (Singh et al. 2018). Similarly, inoculation of siderophore-producing *Arthrobacter sulfonivorans* DS-68 and *Enterococcus hirae* DS-163 endophytes into soils with lower Fe content resulted in an increase in the average number of root tips and root surface area by 1.6 and 2 folds over the control, respectively, according to another study (Singh et al. 2017). However, in the presence of a high Fe concentration in the soil, the average number of root tips and root surface area increased by 1.2 and 1.5 times, respectively, compared to the control. Fortification of wheat grains in this way was made easier by the enlarged root parameters. They then inoculated wheat with *Bacillus subtilis* DS-178 and *Arthrobacter* sp. DS-179 and promising Zn-solubilizers and siderophore-producing endophytes (*Enterococcus hirae* DS-163 and *Arthrobacter sulfonivorans* DS-68) enhanced yield and micronutrient status (Singh et al. 2017).



## Endophytic biofortification

Mycorrhizal fungus inoculation has a beneficial effect on the availability of micronutrients in soil via siderophore synthesis, rhizospheric acidification, and hyphal transfer of nutrients through the external mycelium. Additionally, increased grain Zn concentrations were detected after *Rhizophagus irregularis* colonisation under Zn-deficient circumstances through overexpression of *HvZIP13* (Watts-Williams and Cavagnaro 2018). The combined use of mycorrhizal fungi (MF) and plant growth-promoting bacteria (PGPB) resulted in improved yield-related measures, and increased Zn and Fe grain content in wheat compared to the control group. The PGPB increased the number of metabolic processes contributing to plant growth indices, including photosynthesis. However, AM is involved in several functions, including effective micronutrient absorption in plant tissues and encouragement of plant development through the production of signalling molecules and various other chemicals that alter agricultural plants' root shape and architecture. Along with PGPB and AM fungi, this combination improves stress tolerance, weed management, and better protection against pathogens (Yadav et al. 2020). According to previous studies, endophyte inoculation considerably lowered the phytic acid content in wheat grains by roughly 26% over the RDF, resulting in a rise in Fe and Zn concentrations (Singh et al. 2017).

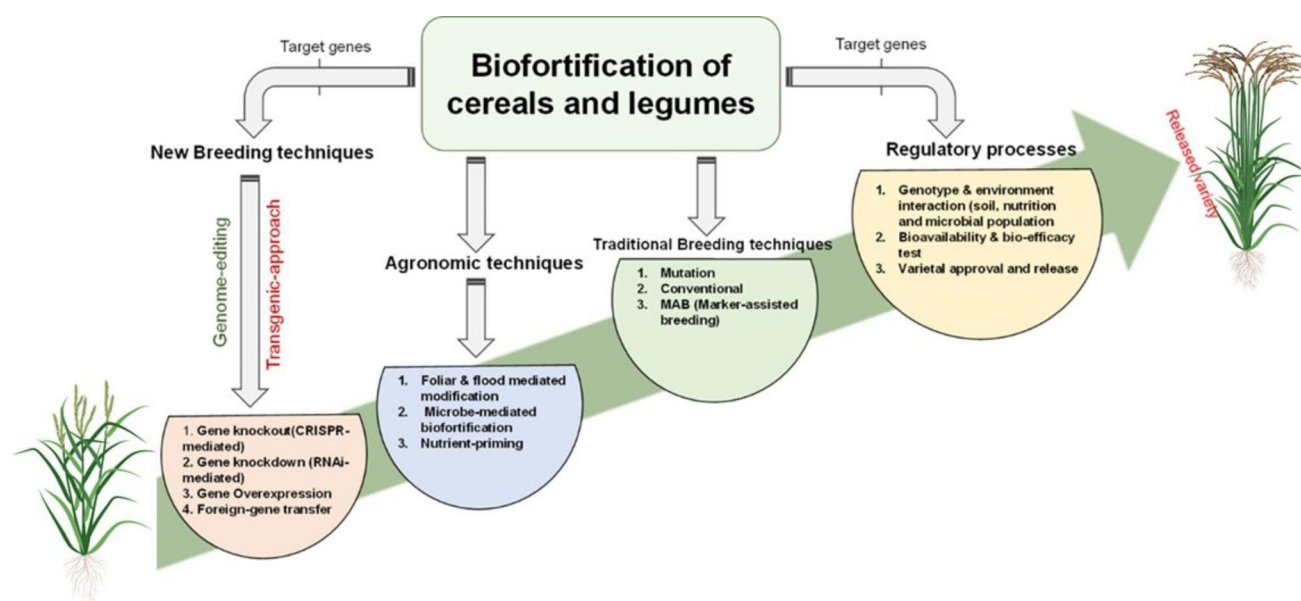
## New breeding technique (Genetic and genome engineering)

New breeding techniques are best strategy of biofortification. This strategy is based on group techniques where Overexpression of genes, Gene transfer, Gene silencing, Gene knockout is performed for biofortification of crops. Knocking out of genes is generally performed to stop the biosynthesis of anti-nutrient compounds. There are various anti-nutrient compounds which are involved in various pathways to restrict the bioavailability of essential micronutrients. These are lectins, phytic acid, saponins, pathogens, protease inhibitors,  $\alpha$ -amylase inhibitors, and tannins. Hence, this results in mal-nutrient crops. Genes involved in the biosynthesis of anti-nutrients could be repressed through RNAi for reduced accumulation of these compounds. Tissue specific expression or overexpression of genes responsible for micronutrient accumulation in plants are the best way for the bioavailability of biofortification in transgenic crops. To improve the nutritional quality and quantity of micronutrients, gene transfer methods of biofortification is generally used. Different genes involved in the biosynthesis of provitamin A (*CrtB*), iron homeostasis (*Fer1-A*), and flavonoid production (*CI*) have been used for biofortification through

various new breeding techniques and metabolic engineering (Shahzad et al., 2021; Blancquaert et al., 2017). These genes are transferred across the various crop species for enhancement of the nutritional supports. These genes help to biofortify the crops for better production of a micronutrient rich diet. In this way we can easily control the malnutrition and disease associated with hidden hunger. Previously, this technology has certain limitations, i.e., laborious, expensive, time-consuming, and regulatory issues but after historical and visionary decision of Govt. of India, SDN1 and few SDN 2 categories are now relaxed for tough regulation steps. It will become easy to develop, produce and transfer the biofortified crops in various cultivars.

## Breeding for genetic biofortification

Plant breeding methods generate crops with greater micronutrient content, moderate to low levels of anti-nutrients, and enhanced amounts of compounds that encourage nutrient absorption via genetic biofortification (Bouis 2003). Breeding techniques can significantly boost micronutrient density by examining genetic diversity to generate nutrient-dense crop variants (Fig. 2). As a result of its long-term viability and lack of regulatory and political restrictions, biofortification through breeding seems to be the best method for biofortification (Saltzman et al. 2017). However, it is necessary to first establish a target micronutrient level for each crop before developing an agriculture crop genetic biofortification programme. Available genetic variability and information about genes that control the absorption of the element by roots, translocation to shoots, mobilization in different vegetative parts, and deposition of the element in edible parts in utilizable forms are among the factors that affect genetic biofortification (Bouis and Welch 2010). Aside from that, various environmental and cultural variables may influence micronutrient accumulation and dietary factors that influence mineral absorption and use by consumers (Bouis and Welch 2010). Breeding has been used to generate most biofortified goods in Asia, Africa, and Latin America, while new methods are being developed to manufacture biofortified products (Garcia-Casal et al. 2017). Biofortified food crops with high nutritional content are grown and marketed by HarvestPlus2 in collaboration with many CGIAR and National Agricultural Research Centers in Africa, Asia, and Latin America. Several biofortified crops, including yellow cassava, orange flesh sweet potato with high levels of  $\beta$ -carotene (over 200 mg/g), iron beans (50–70% more Fe content), orange maize, iron pearl millet, zinc rice, and zinc wheat developed through breeding, have been officially released in more than 30 countries and are in the testing stage in more than 50 countries, according to the HarvestPlus Annual Report (2015). Numerous studies have



**Fig. 2** Biofortification of cereals and legumes through different methodologies. These are broadly classified into three types, i.e., agronomic techniques, new breeding methods, and traditional breeding

approaches. The regulatory process is the most essential stage in the release of a crop variety. Figure created with BioRender (biorender.com)

shown that these newly released biofortified cultivars effectively reduce micronutrient deficiencies in target groups (De Moura et al. 2014; Finkelstein et al. 2017).

In contrast to macronutrients, micronutrients account for a relatively small proportion of the overall weight of a grain crop; as a result, precision measurement is essential for efficiently measuring genetic variation to breed stable and high element concentrations. However, approaches for finding nutrient-dense genotypes that are rapid, precise, and economical have yet to be discovered. Pre-breeding procedures may be employed to create the parent for genetic biofortification if wild relatives are confirmed to be a source of micronutrients.

It is difficult to obtain genetic variation for the goal nutritional trait via selection and transgressive segregation of heterosis (Bouis and Welch 2010). It is necessary to have a basic grasp of genetics to successfully design a breeding programme and pick the appropriate parental line and breeding procedure. Furthermore, a greater understanding of the relationship between nutritional attributes and yield and yield-related traits will assist in the selection of nutrient-rich lines that produce higher yields while still exhibiting desirable characteristics (Bouis and Welch 2010). If, for example, there is a positive association between the Fe and Zn content in the seeds of common bean, peanut, mung bean, wheat, pearl millet, maize, and sesame, this implies the possibility of simultaneously improving these two traits (Pixley et al. 2011; Velu et al. 2012). A substantial negative correlation between Zn and grain yield across sites was identified in the case of chickpeas. In contrast, a significant negative

correlation between Fe and grain yield was detected at one location in the case of chickpea (Diapari et al. 2014).

### Genetic Architecture for the Accumulation of Nutrients

Even though genetic biofortification initiatives have been somewhat successful in addressing the problem of low nutritional content, they have not been as successful as expected in the long run. As a result, it's critical to use genomics ability to speed up the creation of nutrient-dense enhanced cultivars. The details of key crop genetic and genomic resources have been thoroughly studied (Bevan et al. 2017; Varshney et al. 2018; Roorkiwal et al. 2020). Recent advances in next-generation sequencing (NGS) technology have resulted in a dramatic decrease in the cost of sequencing agronomically important crops, allowing the use of NGS-based approaches for allele mining, candidate gene discovery, and high-resolution genetic mapping. Despite cost-effective genotyping systems for applying genomics-assisted breeding (GAB) in agronomically important crops, the expense of high throughput and accurate nutritional estimates remains a significant problem. The plant genome sequencing provides a chance to examine and comprehend the process for functional characterization of genes involved in nutrient absorption and mobilization.

## Identification of QTLs/Genes

Linking genetic data with nutritional content data is a sophisticated and accurate way of identifying quantitative trait loci (QTLs) related to a characteristic of interest, and it is becoming more popular. Nutritional profiling of a broad group of genotypically defined germplasm (core collection, mini-core collection, reference set, composite set) may be used to establish a relationship between genetic data and global mineral nutrition (Ghandilyan et al. 2009; Norton et al. 2010). QTL mapping has become more popular for linking genetic variation to phenotypic variation and as a dependable technique for gene identification. QTL mapping may lead to the isolation or cloning of genetic markers (i.e., related regions or linked regions) that can be used to identify and analyse the genes in question. Many crops have had their genome regions that influence nutritional content/concentration mapped at the molecular level. Studies on understanding genes and processes to enhance seed nutritional composition by discovering QTLs were confined to a few nutrients in crop plants to improve seed nutritional composition. Seed element concentration QTLs have been found in *Lotus Japonicus*, *Medicago truncatula*, common bean, soybean, chickpea, and lentil, for example (Blair et al. 2010, 2011; Casañas et al. 2013; Zhang et al. 2009; Jegadeesan et al. 2010; Ramamurthy et al. 2014; Sab et al. 2020; Aldemir et al. 2017). The majority of research to date to map and tag the gene(s)/QTL(s) determining micronutrient status in crops has identified a quantitative mode of inheritance (Blair et al. 2010). Two genes (*PvIRT1* and *PvIRT2*) on chromosome-3 and two genes (*PvbZIP2* and *PvbZIP3*) on chromosome-11, for example, were linked with QTLs for Fe and Zn in *Proteus vulgaris* (Jiang et al. 2008).

Along with traditional bi-parental mapping populations, attempts have been undertaken to strengthen the available genetic variation for nutritional components via genome-wide association studies (GWAS). This strategy uncovered markers related to various critical nutritional parameters in common bean and chickpea varieties (unpublished) (Katuramu et al. 2018; Caproni et al. 2020). Following confirmation, these newly discovered genes/QTLs may be used to generate nutrient-dense crops high in protein.

## Biofortification via genome engineering

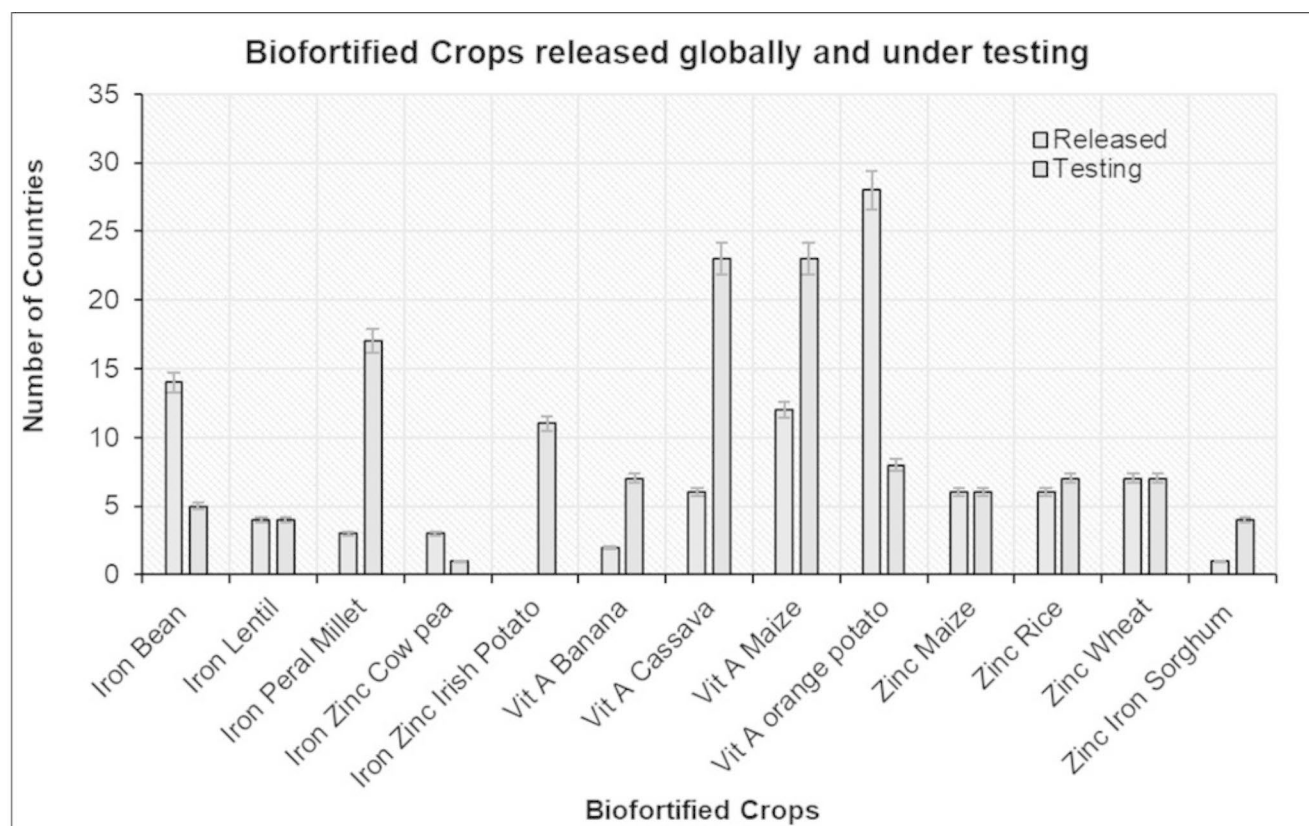
In the absence of natural diversity in the target crop's gene pool, transgenic techniques are required and preferable over breeding (Al-Babili and Beyer 2005). "Golden Rice" is the greatest example of genome engineering for a nutritional attribute since an advanced transgenic line with 37 mg/g carotenoid was generated (Al-Babili and Beyer 2005). Despite the positive side of transgenics, some researchers

have raised concerns about allergies or intolerances caused by bioengineered or genetically modified crops, the environmental side effects, and the loss of biodiversity caused by these crops (Maghari and Ardekani 2011; Raman 2017). In agronomically important crops, recent advancements in molecular biology have drastically altered mutagenesis platforms for more focused and precise DNA modifications using transcription activator-like effector nucleases (TALENs), zinc-finger nucleases (ZFNs), and CRISPR-associated protein (Cas9) (Haun et al. 2014; Michno et al. 2015; Tang et al. 2016). Innovative technologies such as TALENs, ZFNs, RNA interference (RNAi), and CRISPR/Cas9 should be used to enhance nutritional characteristics in many crops plant.

RNAi seems to have an advantage over other genetic engineering methods. It is a sophisticated, particular gene silencing technique and a highly potent breakthrough that can help build nutritionally rich and antinutrient low crops (Tang and Galili 2004). In the past, RNAi technology has been utilised to lower the amount of BOAA in grass pea, reduce the quantity of Arah2, an allergy, by 25% in crude peanut extract, and generate peanut oils that include unique combinations of oleic acid. RNAi has also been utilised to develop common bean lines resistant to the *Golden Mosaic Virus*. (Dodo et al. 2008; Bonfim et al. 2007). Furthermore, the RNAi-mediated suppression of the SACPD gene has boosted soybean resistance to various diseases (Jiang et al. 2009). Because of lower lignin concentrations, RNAi treatments targeting genes involved in lignin synthesis improved soybean resistance to *Sclerotinia sclerotiorum* (Peltier et al. 2009). Additionally, the RNAi method has aided in enhancing oleic acid in soybean. Apart from legumes, it has improved maize, wheat, rice, cotton, jute, and tomato (Kusaba et al. 2003; Davuluri et al. 2005).

## Biofortified crops released or testing by countries worldwide

In a global scenario, most of the countries are working to fight with hidden hunger using various techniques of biofortification. Figure 3 indicates that in which countries biofortified crop varieties had been released or were in testing by the end of the year 2021. Various crop varieties covered have been promoted by HarvestPlus or the International Potato Center, both of which are part of the CGIAR global research partnership for a food secure future. These biofortified varieties have been developed through conventional crop breeding for everyone in Africa, Asia, and Latin America. HarvestPlus promoted biofortified crops which include vitamin A cassava, maize, and sweet potato; iron beans and pearl millet; and zinc rice, maize, and wheat. Many experts in nutrition and agriculture work together to develop these



**Fig. 3** Status of biofortified crops released or under testing by countries worldwide. More than 14 crop varieties released in 92 countries worldwide. Total 128 countries are still working on trails of these all 14 biofortified crops (<https://biocropshplustst.ciat.cgiar.org/>)

biofortified crops. They ensure that how these crops can develop high levels of nutrients to improve nutrition and health. They taste good and meet cooking requirements, and that they have agronomic traits farmers demand such as resistance to insects, diseases, and drought tolerance (Arun and Birol, 2020).

More than 48 million people in smallholder farming households are benefiting from conventionally bred biofortified crops facilitated by HarvestPlus. These crops improve nutrition and health status when eaten regularly. Most of the countries around the world now ready to adopt biofortification as part of their crop improvement programs using a wider strategy to combat micronutrient deficiencies (Fig. 3). HarvestPlus now anticipates that by 2030, around 1 billion people will be benefitting from biofortified foods.

### Biofortified crops released or testing; an indian scenario

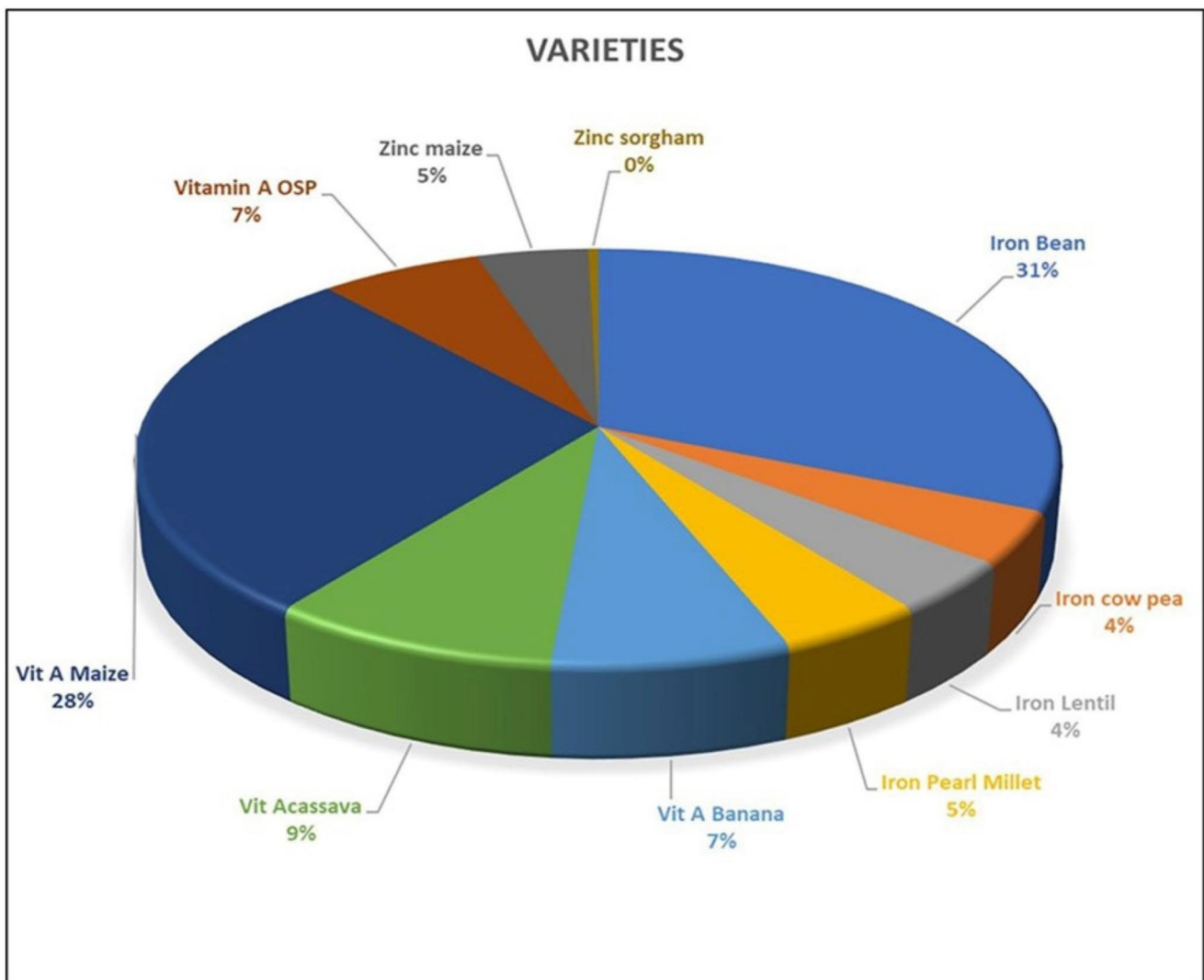
Govt of India have a defined program of biofortification through Indian Council of Agricultural Research for the nutritional biofortification of the staples crops and initiated many programmes in different crops. This body has initiated and recognized these programmes in leadership of Dr.

Trilochan Mohapatra, Director General, ICAR and Secretary DARE. National Agricultural Research System including ICAR and State Agricultural Universities (SAUs) made a significant progress in this area to develop of many biofortified varieties of cereals, pulses, oilseeds, vegetables and fruits along with staples. In the last 3–4 years these developments have published in the form of a report entitled, “Biofortified Varieties: Sustainable Way to Alleviate Malnutrition”. This makes awareness among the stakeholders and consumers, through India and neighbouring countries. The details of released and tested varieties are summarized in Fig. 4.

### Conclusion

Mineral micronutrient absorption and usage efficiency in plants are currently neglected traits in crop breeding projects, despite the fact that they have the potential to increase both the sustainability of agricultural yields and the quality of food and feed. Micronutrient supplementation and fortification procedures are generally advised strategies for alleviating micronutrient deficiency concerns in the human population and livestock. Efforts are being undertaken





**Fig. 4** Biofortified crops and their different varieties released by different countries. A total of 245 biofortified crop varieties has released by various countries for ten different crops (<https://biocropshplustst.ciat.cgiar.org/>)

worldwide to introduce nutrient-rich superfoods like staple grains and millets into mainstream agriculture. Although great progress has been made in producing transgenics for improved Fe and Zn nutrition in crop plants, further research into the holistic genetic circuitry of Fe and Zn intake, transport, and storage in grains is required.

Micronutrient deficiency is projected to rise as the population of developed nations' shifts toward more plant-based diets, and breeding food plants for high mineral micronutrient content looks to be a sustainable solution. Furthermore, breeding for mineral micronutrient efficiency will enable long-term crop production, which is necessary for light of global change. In this respect, it is required to stimulate research and innovation in this subject to protect the well-being of human populations while also maintaining the competitiveness of the agri-food business. As a first step in developing lasting solutions to micronutrient deficiency in

human people, it is vital to understand the molecular and physiological processes underlying micronutrient homeostasis. There has been a plethora of information on the genes and promoters for target features in agronomically essential crop plants. It has been feasible to generate transgenic lines with improved micronutrients. Future studies should try to combine multiple genetic metabolic pathways to boost a variety of nutrients in a specific crop at the same time. Such genetically fortified crops for micronutrients may aid in the alleviation of nutritional issues.

**Author Contribution** "All authors contributed to the study conception and design. Material preparation, data collection and analysis were performed by Sayanti Mandal, Santosh Kumar Gupta, Mimosa Ghorai, Uttpal Anand and Samapika Nandy. The first draft of the manuscript was written by Protha Biswas, Manoj Tukaram Patil, Manoj Kumar, Radha and Abilash Valsala Gopalakrishnan. Figures were prepared by Md. Habibur Rahman, Dorairaj Arvind Prasanth, Abhijit Bhagwan Mane, Niraj Kumar Jha and Saurabh Kumar Jha. Tables were prepared



and the primary draft was revised by Milan Kumar Lal and Rahul Kumar Tiwari. The conceptualization, guidance and resources were provided by Abhijit Dey. All authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.”

**Funding** “The authors declare that no funds, grants, or other support were received during the preparation of this manuscript.”

**Data Availability** “Since it is a review article, no datasets generated during and/or analysed during the current study; all are incorporated in the manuscript.”

## Declarations

**Competing interests** The authors have no relevant financial or non-financial interests to disclose.

## References

- Al-Babili, S., and Beyer, P. (2005). Golden Rice – five yrs. On the road—five yrs. To go? *Trends Plant Sci.* 10, 565–573.
- Aldemir, S., Ates, D., Temel, H. Y., Yagmur, B., Alsaleh, A., Kahriman, A., et al. (2017). QTLs for iron concentration in seeds of the cultivated lentil (*Lens culinaris* Medic.) Via genotyping by sequencing. *Turk J. Agric.* 41, 243–255. doi: <https://doi.org/10.3906/tar-1610-33>
- Andresen E, Peiter E, Küpper H. (2018) Trace metal metabolism in plants. *Journal of Experimental Botany* 69, 909–954.
- Aslam M, Aslam A, Sheraz M, Ali B, Ulhassan Z, Najeeb U, Zhou W, Gill RA. (2021) Lead toxicity in Cereals: mechanistic insight into toxicity, Mode of Action, and management. *Front Plant Sci.* 4;11:587785. doi: <https://doi.org/10.3389/fpls.2020.587785>.
- Balafrej H, Bogusz D, Triqui ZA, Guedira A, Bendaou N, Smouni A, Fahr M. (2020) Zinc Hyperaccumulation in Plants: A Review. *Plants (Basel)*. 29;9(5):562. doi: <https://doi.org/10.3390/plants9050562>
- Bevan, M. W., Uauy, C., Wulff, B. B., Zhou, J., Krasileva, K., and Clark, M. D. (2017). Genomic innovation for crop improvement. *Nature* 543, 346–354.
- Bhatt, P.; Verma, A.; Verma, S.; Anwar, M.S.; Prasher, P.; Mudila, H.; Chen, S. (2020) Understanding phytomicrobiome: a potential reservoir for better crop management. *Sustainability*, 12, 5446.
- Bhupalraj, G.; Patnaik, M.C.; Khadke, K.M. (2002) Molybdenum status in soils of Andhra Pradesh. *AICRP Micro Second. Nutr. Soils Plants Pradesh*, 36, 1–87.
- Blair, M. W., Astudillo, C., Rengifo, J., Beebe, S. E., and Graham, R. (2011). QTL for seed iron and zinc concentrations in a recombinant inbred line population of Andean common beans (*Phaseolus vulgaris* L.). *Theor. Appl. Genet.* 122, 511–523. doi: <https://doi.org/10.1007/s00122-010-1465-8>
- Blair, M. W., Knewton, S. J. B., Astudillo, C., Li, C. M., Fernandez, A. C., and Grusak, M. (2010). Variation and inheritance of iron reductase activity in the roots of common bean (*Phaseolus vulgaris* L.) and association with seed iron accumulation QTL. *BMC Plant Biol.* 10:215. doi: <https://doi.org/10.1186/1471-2229-10-215>
- Blancquaert D, De Steur H, Gellynck X, Van Der Straeten D. (2017) Metabolic engineering of micronutrients in crop plants. *Ann N Y Acad Sci.* 1390(1):59–73. doi: <https://doi.org/10.1111/nyas.13274>.
- Bünemann, E. K., Bongiorno, G., Bai, Z. G., Creamer, R. E., De Deyn, G., de Goede, R., et al. (2018). Soil quality—a critical review. *Soil Biol. Biochem.* 120, 105–125. doi: <https://doi.org/10.1016/j.soilbio.2018.01.030>
- Bonfim, K., Faria, J. C., Nogueira, E. O., Mendes, E. A., and Aragão, F. J. (2007). RNAi-mediated resistance to Bean golden mosaic virus in genetically engineered common bean (*Phaseolus vulgaris*). *Mol. Plant Microbe Interact.* 20, 717–726. doi: <https://doi.org/10.1094/mpmi-20-6-0717>
- Bouis, H. E. (2003). Micronutrient fortification of plants through plant breeding: can it improve nutrition in man at low cost? *Proc. Nutr. Soc.* 62, 403–411. doi: <https://doi.org/10.1079/pns2003262>
- Bouis, H. E., and Welch, R. M. (2010). Biofortification - A sustainable agricultural strategy for reducing micronutrient malnutrition in the global south. *Crop Sci.* 50, S20–S32.
- Braun DM, Wang L, Ruan YL (2014) Understanding and manipulating sucrose phloem loading, unloading, metabolism, and signalling to enhance crop yield and food security. *J Exp Bot* 65:1713–1735. doi: <https://doi.org/10.1093/jxb/ert416>
- Brevik, E.C.; Slaughter, L.; Singh, B.R.; Steffan, J.J.; Collier, D.; Barnhart, P.; Pereira, P. (2020) Soil and human health: current status and future needs. *Air Soil Water Res.*, 13, 1–23.
- Briat JF, Dubos C, Gaymard F (2015 Jan) Iron nutrition, biomass production, and plant product quality. *Trends Plant Sci* 20(1):33–40. doi: <https://doi.org/10.1016/j.tplants.2014.07.005>. Epub 2014 Aug 18. PMID: 25153038.
- Brueck, H.; Lammel, J. (2016) Impact of fertilizer N application on the grey water footprint of winter wheat in a NW-European temperate climate. *Water*, 8, 356.
- Cakmak, I.; Kutman, U.B. (2017) Agronomic biofortification of cereals with zinc: a review. *Eur. J. Soil Sci.* 69, 172–180.
- Cakmak I, Pfeiffer WH, McClafferty B. (2010) Biofortification of durum wheat with zinc and iron. *Cereal Chemistry Journal* 87, 10–20.
- Callens, T.; del Castello, R.; Baratelli, M.; Xipsiti, M.; Navarro, D.K. (2019) The International Year of Pulses; Final Report; FAO: Rome, Italy, p. 40.
- Caproni, L., Raggi, L., Talsma, E. F., Wenzl, P., and Negri, V. (2020). European landrace diversity for common bean biofortification: a genome-wide association study. *Sci. Rep.* 10:19775. doi: <https://doi.org/10.1038/s41598-020-76417-3>
- Casañas, F., Pérez-Vega, E., Almirall, A., Plans, M., Sabaté, J., and Ferreira, J. J. (2013). Mapping of QTL associated with seed chemical content in a RIL population of common bean (*Phaseolus vulgaris* L.). *Euphytica* 192, 279–288. doi: <https://doi.org/10.1007/s10681-013-0880-8>
- Chaiwong, N., Bouain, N., Prom-u-thai, C., and Rouached, H. (2020) Interplay between silicon and iron signalling pathways to regulate silicon transporter Lsi1 expression in rice. *Front. Plant Sci.* 11:1065. doi: <https://doi.org/10.3389/fpls.2020.01065>
- Chen Q, Soulay F, Saudemont B, Elmayan T, Marmagne A, Masclaux-Daubresse C. (2019) Overexpression of *ATG8* in Arabidopsis stimulates autophagic activity and increases nitrogen remobilization efficiency and grain filling. *Plant and Cell Physiology* 60, 343–352.
- Clemens S. (2019) Metal ligands in micronutrient acquisition and homeostasis. *Plant, Cell & Environment* 42, 2902–29
- Cogger, C.; Brown, S. (2016) Soil formation and nutrient cycling. In *Sowing Seeds in the City*; Brown, S., McIvor, K., Hodges, S.E., Eds.; Springer: Dordrecht, The Netherlands.
- Connorton, J.M.; Balk, J. (2019) Iron biofortification of staple crops: Lessons and challenges in plant genetics. *Plant Cell Physiol.* 60, 1447–1456.
- Connorton JM, Balk J, Rodríguez-Celma J. (2017) Iron homeostasis in plants – a brief overview. *Metallomics* 9, 813–823.
- Davuluri, G. R., van Tuinen, A., Fraser, P. D., Manfredonia, A., Newman, R., Burgess, D., et al. (2005). Fruit-specific RNAi-mediated suppression of DET1 enhances carotenoid and flavonoid content in tomatoes. *Nat. Biotechnol.* 23, 890–895. doi: <https://doi.org/10.1038/nbt1108>

- De Moura, F. F., Palmer, A. C., Finkelstein, J. L., Haas, J. D., Murray-Kolb, L. E., and Wenger, M. J. (2014). Are biofortified staple food crops improving vitamin a and iron status in women and children? New evidence from efficacy trials. *Adv. Nutr.* 5, 568–570. doi: <https://doi.org/10.3945/an.114.006627>
- Diapari, M., Sindhu, A., Bett, K., Deokar, A., Warkentin, T. D., and Tar'an, B. (2014). Genetic diversity and association mapping of iron and zinc concentrations in chickpea (*Cicer arietinum* L.). *Genome* 57, 459–468. doi: <https://doi.org/10.1139/gen-2014-0108>
- Dodo, H. W., Konan, K. N., Chen, F. C., Egnin, M., and Viquez, O. M. (2008). Alleviating peanut allergy using genetic engineering: the silencing of the immunodominant allergen Ara h 2 leads to its significant reduction and a decrease in peanut allergenicity. *Plant Biotechnol. J.* 6, 135–145. doi: <https://doi.org/10.1111/j.1467-7652.2007.00292.x>
- Dong J, Piñeros MA, Li X, Yang H, Liu Y, Murphy AS, Kochian LV, Liu D. (2017) An Arabidopsis ABC transporter mediates phosphate deficiency-induced remodeling of root architecture by modulating iron homeostasis in roots. *Molecular Plant* 10, 244–259.
- Fendrihan, S.; Pop, C.E. (2021) Biotechnological potential of associated microorganism. *Rom. Biotechnol. Lett.*, 26, 2700–2706.
- Finkelstein, J. L., Haas, J. D., and Mehta, S. (2017). Iron-biofortified staple food crops for improving iron status: a review of the current evidence. *Curr. Opin. Biotechnol.* 44, 138–145. doi: <https://doi.org/10.1016/j.copbio.2017.01.003>
- Flis P, Ouerdane L, Grillet L, Curie C, Mari S, Lobinski R. (2016) Inventory of metal complexes circulating in plant fluids: a reliable method based on HPLC coupled with dual elemental and high-resolution molecular mass spectrometric detection. *New Phytologist* 211, 1129–1141.
- Garcia-Casal, M. N., Peña-Rosas, J. P., Giyose, B., De Steur, H., and Van Der Straeten, D. (2017). Staple crops biofortified with increased vitamins and minerals: considerations for a public health strategy. *Ann. N.Y. Acad. Sci.* 1379, 38–47.
- Ghandilyan, A., Barboza, L., Tisné, S., Granier, C., Reymond, M., Koornneef, M., et al. (2009). Genetic analysis identifies quantitative trait loci controlling rosette mineral concentrations in *Arabidopsis thaliana* under drought. *New Phytol.* 184, 180–192. doi: <https://doi.org/10.1111/j.1469-8137.2009.02953.x>
- González-Guerrero, M., Escudero, V., Saéz, Á., and Tejada-Jiménez, M. (2016) Transition metal transport in plants and associated endosymbionts: arbuscular mycorrhizal fungi and rhizobia. *Front. Plant Sci.* 7:1088.
- Grillet L, Ouerdane L, Flis P, Hoang MTT, Isaure M-P, Lobinski R, Curie C, Mari S. (2014) Ascorbate efflux as a new strategy for iron reduction and transport in plants. *Journal of Biological Chemistry* 289, 2515–2525.
- Górniak, W.; Cholewińska, P.; Konkol, D. (2018) Feed additives produced on the basis of organic forms of micronutrients as a means of biofortification of food of animal origin. *J. Chem.* 8084127.
- Gruber, B. D., Giehl, R. F., Friedel, S., and von Wirén, N. (2013) Plasticity of the Arabidopsis root system under nutrient deficiencies. *Plant Physiol.* 163, 161–179. doi: <https://doi.org/10.1104/pp.113.218453>
- Hanikenne M, Esteves SM, Fanara S, Rouached H. (2021) Coordinated homeostasis of essential mineral nutrients: a focus on iron. *Journal of Experimental Botany* 72, 2136–2153.
- HarvestPlus Annual Report (2015). HarvestPlus Annual Report. [https://www.harvestplus.org/knowledge-market/publications?%5B0%5D=field\\_resource\\_type%3A11](https://www.harvestplus.org/knowledge-market/publications?%5B0%5D=field_resource_type%3A11)
- Haun, W., Coffman, A., Clasen, B. M., Demorest, Z. L., Lowy, A., Ray, E., et al. (2014). Improved soybean oil quality by targeted mutagenesis of the fatty acid desaturase 2 gene family. *Plant Biotechnol. J.* 12, 934–940. doi: <https://doi.org/10.1111/pbi.12201>
- Hindt MN, Guerinot ML (2012 Sep) Getting a sense for signals: regulation of the plant iron deficiency response. *Biochim Biophys Acta* 1823(9):1521–30. <https://doi.org/10.1016/j.bbamcr.2012.03.010>
- Huang, S.; Wang, P.; Yamaji, N.; Ma, J.F. (2020) Plant nutrition for human nutrition: hints from rice research and future perspectives. *Mol. Plant*, 13, 825–835.
- Ishikawa S, Ishimaru Y, Igura M, Kuramata M, Abe T, Senoura T, Hase Y, Arai T, Nishizawa NK, Nakanishi H. (2012) Ion-beam irradiation, gene identification, and marker-assisted breeding in the development of low-cadmium rice. *PNAS, USA* 109, 19166–19171.
- Jaiswal, D.K., Krishna, R., Chouhan, G.K. et al (2022) Bio-fortification of minerals in crops: current scenario and future prospects for sustainable agriculture and human health. *Plant Growth Regul* 98, 5–22. <https://doi.org/10.1007/s10725-022-00847-4>
- Jegadeesan, S., Yu, K., Poysa, V., Gawalko, E., Morrison, M. J., Shi, C., et al. (2010). Mapping and validation of simple sequence repeat markers linked to a major gene controlling seed cadmium accumulation in soybean [*Glycine max* (L.) Merr.]. *Theor. Appl. Genet.* 121, 283–294. doi: <https://doi.org/10.1007/s00122-010-1309-6>
- Jeong, J., Merkovich, A., Clyne, M., and Connolly, E. L. (2017) Directing iron transport in dicots: regulation of iron acquisition and translocation. *Curr. Opin. Plant Biol.* 39, 106–113. doi: <https://doi.org/10.1016/j.pbi.2017.06.014>
- Jiang, D., Zhao, L., and Clapham, D. E. (2009). Genome-wide RNAi screen identifies Letm1 as a mitochondrial  $\text{Ca}_2^+/\text{H}^+$  antiporter. *Science* 326, 144–147. doi: <https://doi.org/10.1126/science.1175145>
- Jiang, W., Struik, P. C., van Keulen, H., Zhao, M., Jin, L. N., and Stomph, T. J. (2008). Does increased zinc uptake enhance grain zinc mass concentration in rice? *Ann. Appl. Biol.* 153, 135–147. doi: <https://doi.org/10.1111/j.1744-7348.2008.00243.x>
- Katuramu D, Hart J, Porch T, Grusak M, Glahn R, Cichy K (2018) Genome-wide association analysis of nutritional composition-related traits and iron bioavailability in cooked dry beans (*Phaseolus vulgaris* L.). *Mol Breed* 38:44. <https://doi.org/10.1007/s11032-018-0798-x>
- Kaur, T.; Rana, K.L.; Kour, D.; Sheikh, I.; Yadav, N.; Kumar, V.; Yadav, A.N.; Dhaliwal, H.S.; Saxena, A.K. (2020) Microbe-mediated biofortification for micronutrients: Present status and future challenges. In *New and Future Developments in Microbial Biotechnology and Bioengineering*; Elsevier: Amsterdam, The Netherlands, pp. 1–17.
- Kawakami Y, Bhullar NK. (2021) Delineating the future of iron biofortification studies in rice: challenges and future perspectives. *Journal of Experimental Botany* 72, 2099–2113.
- Khoshgofarmanesh, A.H., Schulin, R., Chaney, R.L. et al (2010) Micronutrient-efficient genotypes for crop yield and nutritional quality in sustainable agriculture. A review. *Agron. Sustain. Dev.* 30, 83–107. <https://doi.org/10.1051/agro/2009017>
- Kihara, J.; Bolo, P.; Kinyua, M.; Rurinda, J.; Pikki, K. (2020) Micronutrient deficiencies in african soils and the human nutritional nexus: Opportunities with staple crops. *Environ. Geochem. Health*, 42, 3015–3033.
- Kisko M, Bouain N, Safi A, et al. (2018) LPCAT1 controls phosphate homeostasis in a zinc-dependent manner. *eLife* 7, e32077.
- Klikocka H, Marks M. (2018) Sulphur and nitrogen fertilization as a potential means of agronomic biofortification to improve the content and uptake of microelements in spring wheat grain DM. *J Chem.* 2018:9326820. doi: <https://doi.org/10.1155/2018/9326820>
- Kobayashi T, Nozoye T, Nishizawa NK (2019) Iron transport and its regulation in plants. *Free Rad Biol Med* 133:11–20
- Kowalska, G.; Kowalski, R.; Hawlena, J.; Rowiński, R. (2020) Seeds of oilseed rape as an alternative source of protein and mineral. *J. Elementol.*, 25, 513–522.
- Krouk, G., and Kiba, T. (2020) Nitrogen and phosphorus interactions in plants: from agronomic to physiological and molecular insights.

- Curr. Opin. Plant Biol. 57, 1–6. doi: <https://doi.org/10.1016/j.pbi.2020.07.002>
- Krouk, G., Ruffel, S., Gutiérrez, R. A., Gojon, A., Crawford, N. M., Coruzzi, G. M., et al. (2011) A framework integrating plant growth with hormones and nutrients. *Trends Plant Sci.* 16, 178–182. doi: <https://doi.org/10.1016/j.tplants.2011.02.004>
- Kumar, D.; Dhaliwal, S.S.; Naresh, R.K.; Salaria, A. (2018) Agronomic biofortification of paddy through nitrogen, zinc and iron fertilization: a review. *Int. J. Curr. Microbiol. Appl. Sci.*, 7, 2942–2953.
- Kusaba, M., Miyahara, K., Iid, S., Fukuoka, H., Takano, T., Sassa, H., et al. (2003). Low glutelin content1: a dominant mutation that suppresses the glutelin multigene family via RNA silencing in rice. *Plant Cell* 15, 1455–1467. doi: <https://doi.org/10.1105/tpc.011452>
- Kutman UB, Yildiz B, Cakmak I. (2011) Effect of nitrogen on uptake, remobilization and partitioning of zinc and iron throughout the development of durum wheat. *Plant and Soil* 342, 149–164.
- Lean, M.E. (2019) Principles of human nutrition. *Medicine*, 47, 140–144.
- Lee S, Persson DP, Hansen TH, et al. (2011) Bio-available zinc in rice seeds is increased by activation tagging of nicotianamine synthase: activation and overexpression of OsNAS2. *Plant Biotechnology Journal* 9, 865–73.
- Lešková, A.; Giehl, R.F.H.; Hartmann, A.; Fargašová, A.; von Wirén, N. (2017) Heavy metals induce iron deficiency responses at different hierarchic and regulatory levels. *Plant Physiol.*, 174, 1648–1668.
- Ligowe, I.S.; Bailey, E.H.; Young, S.D.; Ander, E.L.; Kabambe, V.; Chilimba, A.D.; Lark, R.M.; Nalivata, P.C. (2021) Agronomic iodine biofortification of leafy vegetables grown in Vertisols, Oxisols and Alfisols. *Environ. Geochem. Health* 43, 361–374.
- Liu, T. Y., Chang, C. Y., and Chiou, T. J. (2009) The long-distance signalling of mineral macronutrients. *Curr. Opin. Plant Biol.* 12, 312–319. doi: [10.1016/j.pbi.2009.04.00](https://doi.org/10.1016/j.pbi.2009.04.00)
- Maghari, B. M., and Ardekani, A. M. (2011). Genetically modified foods and social concerns. *Avicenna J. Med. Biotechnol.* 3, 109–117.
- Maillard A, Etienne P, Diquélou S, Trouverie J, Billard V, Yvin J-C, Ourry A. (2016a) Nutrient deficiencies modify the ionic composition of plant tissues: a focus on cross-talk between molybdenum and other nutrients in Brassica napus. *Journal of Experimental Botany* 67, 5631–5641.
- Maillard A, Sorin E, Etienne P, et al. (2016b) Non-specific root transport of nutrient gives access to an early nutritional indicator: the case of sulfate and molybdate. *PLoS One* 11, e0166910.
- Mari S, Bailly C, Thomine S. (2020) Handing off iron to the next generation: how does it get into seeds and what for? *Biochemical Journal* 477, 259–274.
- Marschner P, ed. (2012) Marschner's mineral nutrition of higher plants, 3rd edn. London: Elsevier/Academic Press
- Ma X, Geng Q, Zhang H, Bian C, Chen HYH, Jiang D, Xu X. (2021) Global negative effects of nutrient enrichment on arbuscular mycorrhizal fungi, plant diversity and ecosystem multifunctionality. *New Phytologist* 229, 2957–2969.
- Michno, J. M., Wang, X., Liu, J., Curtin, S. J., Kono, T. J., and Stupar, R. M. (2015). CRISPR/Cas mutagenesis of soybean and Medicago truncatula using a new web-tool and a modified Cas9 enzyme. *GM Crops Food* 6, 243–252. doi: <https://doi.org/10.1080/21645698.2015.1106063>
- Morales F, Ancin M, Fakhet D, González-Torralba J, Gámez AL, Seminario A, Soba D, Ben Mariem S, Garriga M, Aranjuelo I. (2020) Photosynthetic Metabolism under Stressful Growth Conditions as a Bases for Crop Breeding and Yield Improvement. *Plants* (Basel). 10;9(1):88. doi: <https://doi.org/10.3390/plants9010088>.
- Morales, F.; Pavlovic, A.; Abadía, A.; Abadía, J. (2018) Photosynthesis in poor nutrient soils, in compacted soils, and under drought. In *The Leaf: a platform for performing photosynthesis*; Adams, W.W., III, Terashima, I., Eds.; Springer: Berlin/Heidelberg, Germany, pp. 371–399.
- Murgia I, De Gara L, Grusak MA. (2013) Biofortification: how can we exploit plant science and biotechnology to reduce micronutrient deficiencies? *Front Plant Sci.* 6;4:429. doi: <https://doi.org/10.3389/fpls.2013.00429>.
- Mu S, Yamaji N, Sasaki A, et al. (2021) A transporter for delivering zinc to the developing tiller bud and panicle in rice. *The Plant Journal* 105, 786–799.
- Norton, G. J., Deacon, C. M., Xion, L., Huang, S., Meharg, A. A., and Price, A. H. (2010). Genetic mapping of the rice ionome in leaves and grain: identification of QTLs for 17 elements including arsenic, cadmium, iron and selenium. *Plant Soil* 329, 139–153. doi: <https://doi.org/10.1007/s11104-009-0141-8>
- Nuss, E.T.; Tanumihardjo, S.A. (2010) Maize: a paramount staple crop in the context of global nutrition. *Food Soc. Food Saf.*, 9, 417–436.
- Olsen LI, Hansen TH, Larue C, et al. (2016) Mother-plant-mediated pumping of zinc into the developing seed. *Nature Plants* 2, 16036.
- Olsen LI, Palmgren MG. (2014) Many rivers to cross: the journey of zinc from soil to seed. *Frontiers in Plant Science* 5, 30
- Peltier, A. J., Hatfield, R. D., and Grau, C. R. (2009). Soybean stem lignin concentration relates to resistance to Sclerotinia sclerotiorum. *Plant Dis.* 93, 149–154. doi: <https://doi.org/10.1094/pdis-93-2-0149>
- Pilon-Smits EA, Quinn CF, Tapken W, Malagoli M, Schiavon M (2009 Jun) Physiological functions of beneficial elements. *Curr Opin Plant Biol* 12(3):267–74. <https://doi.org/10.1016/j.pbi.2009.04.009>.
- Piotrowska-Dlugosz, A.; Siwik-Ziomek, A.; Dlugosz, J.; Gozdowski, D. (2017) Spatio-temporal variability of soil sulfur content and arylsulfatase activity at a conventionally managed arable field. *Geoderma*, 295, 107–118.
- Pixley, K. V., Palacios-Rojas, N., and Glahn, R. P. (2011). The usefulness of iron bioavailability as a target trait for breeding maize (*Zea mays* L.) with enhanced nutritional value. *Field Crops Res.* 123, 153–160. doi: <https://doi.org/10.1016/j.fcr.2011.05.011>
- Pottier M, Dumont J, Masclaux-Daubresse C, Thomine S. (2019) Autophagy is essential for optimal translocation of iron to seeds in Arabidopsis. *Journal of Experimental Botany* 70, 859–869.
- Pottier M, Oomen R, Picco C, Giraudat J, Scholz-Starke J, Richaud P, Carpaneto A, Thomine S. (2015) Identification of mutations allowing Natural Resistance Associated Macrophage Proteins (NRAMP) to discriminate against cadmium. *The Plant Journal* 83, 625–637
- Praharaj S, Skalicky M, Maitra S, Bhadra P, Shankar T, Brestic M, Hejnak V, Vachova P, Hossain A. (2021) Zinc Biofortification in Food Crops Could Alleviate the Zinc Malnutrition in Human Health. *Molecules*. 9;26(12):3509. doi: <https://doi.org/10.3390/molecules26123509>.
- Raffa CM, Chiampo F, Shanthakumar S. (2021) Remediation of Metal/Metalloid-Polluted soils: a short review. *Applied Sciences*. 11(9):4134. <https://doi.org/10.3390/app11094134>
- Ramamurthy, R. K., Jedlicka, J., Graef, G. L., and Waters, B. M. (2014). Identification of new QTLs for seed mineral, cysteine, and methionine concentrations in soybean [*Glycine max* (L.) Merr.]. *Mol. Breed.* 34, 431–445. doi: <https://doi.org/10.1007/s11032-014-0045-z>
- Raman, R. (2017). The impact of genetically modified (GM) crops in modern agriculture: a review. *GM Crops Food*. 8, 195–208. doi: <https://doi.org/10.1080/21645698.2017.1413522>
- Rebello, C.J.; Greenway, F.L.; Finley, J.W. (2014) Whole grains and pulses: a comparison of the nutritional and health benefits. *J Agric. Food Chem.*, 62, 7029–7049.



- Roorkiwal, M., Bharadwaj, C., Barmukh, R., Dixit, G. P., Thudi, M., Gaur, P. M., et al. (2020). Integrating genomics for chickpea improvement: achievements and opportunities. *Theor. Appl. Genet* 133, 1703–1720. doi: <https://doi.org/10.1007/s00122-020-03584-2>
- Rosa-Sibakov, N.; Poutanen, K.; Micard, V. (2015) How does wheat grain, bran and aleurone structure impact their nutritional and technological properties? *Trends Food Sci. Technol.*, 41, 118–134.
- Sab, S., Loksha, R., Mannur, D. M., Somasekhar, Jadhav, K., Mallikarjuna, B. P., et al. (2020). Genome wide SNP discovery and mapping QTLs for seed iron and zinc concentrations in chickpea (*Cicer arietinum* L.). *Front. Nutr.* 7:559120. doi: <https://doi.org/10.3389/fnut.2020.559120>
- Saeid, A.; Patel, A.; Jastrzębska, M.; Korczyński, M. (2019) Food biofortification. *J. Chem.* 5718426.
- Saini RK, Nile SH, Keum Y-S (2016) Food science and technology for management of iron deficiency in humans: a review. *Trends Food Sci Technol* 53:13–22
- Saltzman, A., Birol, E., Oparinde, A., Andersson, M. S., Asare-Marfo, D., Diressie, M. T., et al. (2017). Availability, production, and consumption of crops biofortified by plant breeding: current evidence and future potential. *Ann. N. Y. Acad. Sci.* 1390, 104–114. doi: <https://doi.org/10.1111/nyas.13314>
- Senovilla M, Abreu I, Escudero V, Cano C, Bago A, Imperial J, González-Guerrero M. (2020) MtCOPT2 is a Cu<sup>+</sup> transporter specifically expressed in *Medicago truncatula* mycorrhizal roots. *Mycorrhiza* 30, 781–788.
- Shahzad R, Jamil S, Ahmad S, Nisar A, Khan S, Amina Z, Kanwal S, Aslam HMU, Gill RA, Zhou W. (2021) Biofortification of Cereals and Pulses using new breeding techniques: current and future perspectives. *Front Nutr.* 7:721728. doi: <https://doi.org/10.3389/fnut.2021.721728>
- Shao JF, Yamaji N, Liu XW, Yokosho K, Shen RF, Ma JF. (2018) Preferential distribution of boron to developing tissues is mediated by the intrinsic protein OsNIP3. *Plant Physiology* 176, 1739–1750.
- Shelef, O., Weisberg, P. J., & Provenza, F. D. (2017). The Value of Native Plants and Local Production in an Era of Global Agriculture. *Frontiers in plant science*, 8, 2069. <https://doi.org/10.3389/fpls.2017.02069>
- Sheraz S, Wan Y, Venter E, Verma SK, Xiong Q, Waites J, Connorton JM, Shewry PR, Moore KL, Balk J. (2021) Subcellular dynamics studies of iron reveal how tissue-specific distribution patterns are established in developing wheat grains. *New Phytologist* 231, 1644–1657.
- Shukla, A.K.; Behera, S.K.; Pakhre, A.; Chaudhari, S.K. (2018) Micronutrients in soils, plants, animals and humans. *Indian J. Fert.* 14, 30–54.
- Shukla AK, Behera SK, Prakash C, Patra AK, Rao CS, Chaudhari SK, Das S, Singh AK, Green A. (2021) Assessing Multi-Micronutrients Deficiency in Agricultural Soils of India. *Sustainability*. 13(16):9136. <https://doi.org/10.3390/su13169136>
- Shukla, A.K.; Behera, S.K.; Satyanarayana, T.; Majumdar, K. (2019) Importance of micronutrients in indian agriculture. *Better Crops South Asia*, 11, 6–10.
- Singh AP, Singh MV, Sakal R, Chaudhary VC (2006) Boron nutrition of crops and soils of Bihar. *Tech Bull* 6:1–80.
- Singh, B.R.; Timsina, Y.N.; Lind, O.C.; Cagno, S.; Janssens, K. (2018) Zinc and iron concentration as affected by nitrogen fertilization and their localization in wheat grain. *Front. Plant Sci.*, 9, 307.
- Singh, D.; Rajawat, M.V.S.; Kaushik, R. (2017) Beneficial role of endophytes in biofortification of Zn in wheat genotypes varying in nutrient use efficiency grown in soils sufficient and deficient in Zn. *Plant Soil*, 416, 107–116.
- Singh, M.V. (2001) Evaluation of micronutrient status in different agroecological zones of India. *Fertil. News*, 46, 25–42.
- Sperotto, R. A., Ricachenevsky, F. K., Williams, L. E., Vasconcelos, M. W., and Menguer, P. K. (2014) From soil to seed: micronutrient movement into and within the plant. *Front. Plant Sci.* 5:438.
- Stanton C, Sanders D, Krämer U, Podar D. (2021) Zinc in plants: integrating homeostasis and biofortification. *Molecular Plant* 15, 65–85.
- Steur, H.D.; Mehta, S.; Gellynck, X.; Finkelstein, J.L. (2017) GM biofortified crops: potential effects on targeting the micronutrient intake gap in human populations. *Curr. Opin. Biotechnol.* 44, 181–188.
- Stoyanova, Z. and Doncheva S. (2002) The effect of zinc supply and succinate treatment on plant growth and mineral uptake in pea plant. *Brazilian Journal of Plant Physiology*. 14:111–116. DOI: <https://doi.org/10.1590/S1677-04202002000200005>.
- Stringlis IA, Yu K, Feussner K, de Jonge R, Van Bentum S, Van Verk MC, Berendsen RL, Bakker PAHM, Feussner I, Pieterse CMJ. (2018) MYB72-dependent coumarin exudation shapes root microbiome assembly to promote plant health. *Proceedings of the National Academy of Sciences, USA* 115, E5213–E5222.
- Szákóvá, J.; Praus, L.; Tremlová, J.; Kulhánek, M.; Tlustoš, P. (2017) Efficiency of foliar selenium application on oilseed rape (*Brassica napus* L.) as influenced by rainfall and soil characteristics. *Arch. Agron. Soil Sci.*, 63, 1240–1254.
- Tang, F., Yang, S., Liu, J., and Zhu, H. (2016). Rj4, a gene controlling nodulation specificity in soybeans, encodes a thaumatin-like protein but not the one previously reported. *Plant Physiol.* 170, 26–32. doi: <https://doi.org/10.1104/pp.15.01661>
- Tang, G., and Galili, G. (2004). Using RNAi to improve plant nutritional value: from mechanism to application. *Trends Biotechnol.* 22, 463–469. doi: 10.1016/j.tibtech.2004.07.009
- Tsai HH, Schmidt W. (2017) Mobilization of iron by plant-borne coumarins. *Trends in Plant Science* 22, 538–548.
- Ueno D, Yamaji N, Kono I, Huang CF, Ando T, Yano M, Ma JF. (2010) Gene limiting cadmium accumulation in rice. *Proceedings of the National Academy of Sciences, USA* 107, 16500–16505.
- Varshney, R. K., Thudi, M., Pandey, M. K., Tardieu, F., Ojiewo, C., Vadez, V., et al. (2018). Accelerating genetic gains in legumes for the development of prosperous smallholder agriculture: integrating genomics, phenotyping, systems modelling and agronomy. *J. Exp. Bot.* 69, 3293–3312. doi: 10.1093/jxb/ery088
- Velu, G., Singh, R. P., Huerta-Espino, J., Peña, R. J., Arun, B., Mahendru-Singh, A., et al. (2012). Performance of biofortified spring wheat genotypes in target environments for grain zinc and iron concentrations. *Field Crops Res.* 137, 261–267. doi: <https://doi.org/10.1016/j.fcr.2012.07.018>
- Venkidasamy, B.; Selvaraj, D.; Nile, A.S.; Ramalingam, S.; Kai, G.; Nile, S.H. (2019) Indian pulses: a review on nutritional, functional and biochemical properties with future perspectives. *Trends Food Sci. Technol.*, 88, 228–242.
- Waters BM, Sankaran RP. Moving micronutrients from the soil to the seeds: genes and physiological processes from a biofortification perspective. *Plant Sci.* 2011 Apr;180(4):562–74. doi: <https://doi.org/10.1016/j.plantsci.2010.12.003>.
- Watts-Williams, S.J.; Cavagnaro, T.R. (2018) Arbuscular mycorrhizal fungi increase grain zinc concentration and modify the expression of root ZIP transporter genes in a modern barley (*Hordeum vulgare*) cultivar. *Plant Sci.*, 274, 163–170.
- Watts-Williams SJ, Smith FA, McLaughlin MJ, Patti AF, Cavagnaro TR. (2015) How important is the mycorrhizal pathway for plant Zn uptake? *Plant and Soil* 390, 157–166.
- Wawrzyńska, A., and Sirko, A. (2014) To control and to be controlled: understanding the Arabidopsis *SLIM1* function in sulphur deficiency through comprehensive investigation of the EIL protein family. *Front. Plant Sci.* 5:575. doi: <https://doi.org/10.3389/fpls.2014.00575>

- Xue, Y., Xia, H., Christie, P., Zhang, Z., Li, L., and Tang, C. (2016) Crop acquisition of phosphorus, iron and zinc from soil in cereal/legume intercropping systems: a critical review. *Ann. Bot.* 117, 363–377. doi: <https://doi.org/10.1093/aob/mcv182>
- Yadav, R.; Ror, P.; Rathore, P.; Ramakrishna, W. (2020) Bacteria from native soil in combination with arbuscular mycorrhizal fungi augment wheat yield and biofortification. *Plant Physiol. Biochem.*, 150, 222–233.
- Yazici MA, Asif M, Tutus Y, Ortas I, Ozturk L, Lambers H, Cakmak I. (2021) Reduced root mycorrhizal colonization as affected by phosphorus fertilization is responsible for high cadmium accumulation in wheat. *Plant and Soil* 468, 19–35.
- Zafar, S.; Li, Y.L.; Li, N.N.; Zhu, K.M.; Tan, X.L. (2019) Recent advances in enhancement of oil content in oilseed crops. *J. Biotechnol.*, 301, 35–44.
- Zhang, B., Chen, P., Shi, A., Hou, A., Ishibashi, T., and Wang, D. (2009). Putative quantitative trait loci associated with calcium content in soybean seed. *J. Hered.* 100, 263–262. doi: <https://doi.org/10.1093/jhered/esn096>
- Zhang J, Zhu Y, Yu L, Yang M, Zou X, Yin C, Lin Y. (2022) Research Advances in Cadmium Uptake, Transport and Resistance in Rice (*Oryza sativa* L.). *Cells*. 6;11(3):569. doi: <https://doi.org/10.3390/cells11030569>.
- Zhao, K., and Wu, Y. (2017) Effects of Zn deficiency and bicarbonate on the growth and photosynthetic characteristics of four plant species. *PLoS One* 12:e0169812. doi: <https://doi.org/10.1371/journal.pone.0189620>

**Publisher's Note** Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.

## Authors and Affiliations

Sayanti Mandal<sup>1</sup> · Santosh Kumar Gupta<sup>2</sup> · Mimosa Ghorai<sup>3,4</sup> · Manoj Tukaram Patil<sup>5</sup> · Protha Biswas<sup>3</sup> · Manoj Kumar<sup>6</sup> · Radha<sup>7</sup> · Abilash Valsala Gopalakrishnan<sup>8</sup> · Vikas Moreshwar Mohture<sup>9</sup> · Md. Habibur Rahman<sup>10</sup> · Dorairaj Arvind Prasanth<sup>4</sup> · Abhijit Bhagwan Mane<sup>11,12</sup> · Niraj Kumar Jha<sup>13,14,15,16</sup> · Saurabh Kumar Jha<sup>13,14,15</sup> · Milan Kumar Lal<sup>17</sup> · Rahul Kumar Tiwari<sup>17</sup> · Abhijit Dey<sup>3</sup>

✉ Abhijit Dey  
abhijit.dbs@presiuniv.ac.in

<sup>1</sup> Institute of Bioinformatics and Biotechnology, Savitribai Phule Pune University, Ganeshkhind Road, Pune, Maharashtra 411007, India

<sup>2</sup> National Institute of Plant Genome Research, New Delhi 110067, India

<sup>3</sup> Department of Life Sciences, Presidency University, 86/1 College Street, Kolkata, West Bengal 700073, India

<sup>4</sup> Department of Microbiology, School of Biosciences, Periyar University, Salem, Tamilnadu 636011, India

<sup>5</sup> Post Graduate Department of Botany, SNJB's KKHA Arts SMGL Commerce and SPJ Science College Chandwad (Nashik) Maharashtra, Nashik, India

<sup>6</sup> Chemical and Biochemical Processing Division, ICAR-Central Institute for Research on Cotton Technology, Mumbai 400019, India

<sup>7</sup> School of Biological and Environmental Sciences, Shoolini University of Biotechnology and Management Sciences, Solan, Himachal Pradesh 173229, India

<sup>8</sup> Department of Biomedical Sciences, School of Bio Sciences and Technology, Vellore Institute of Technology (VIT), Vellore, Tamil Nadu 632014, India

<sup>9</sup> Department of Botany, Rashtrapita Mahatma Gandhi Arts and Science College Nagbhid (M.S.), Nagbhid 441205, India

<sup>10</sup> Organic Health Care Ltd., Dhaka, Bangladesh

<sup>11</sup> Department of Zoology, Dr. Patangrao Kadam Mahavidhyalaya, Ramanandnagar (Burli), Sangli, Affiliated, Maharashtra 416308, India

<sup>12</sup> Shivaji University, Kolhapur, India

<sup>13</sup> Department of Biotechnology, School of Engineering and Technology, Sharda University, Greater Noida, Noida, UP 201310, India

<sup>14</sup> Department of Biotechnology Engineering and Food Technology, Chandigarh University, Mohali 140413, India

<sup>15</sup> Department of Biotechnology, School of Applied & Life Sciences (SALS), Uttarakhand University, Dehradun 248007, India

<sup>16</sup> School of Bioengineering & Biosciences, Lovely Professional University, Phagwara 144411, India

<sup>17</sup> ICAR-Central Potato Research Institute, Shimla, Himachal Pradesh 171001, India



## Terms and Conditions

Springer Nature journal content, brought to you courtesy of Springer Nature Customer Service Center GmbH (“Springer Nature”).

Springer Nature supports a reasonable amount of sharing of research papers by authors, subscribers and authorised users (“Users”), for small-scale personal, non-commercial use provided that all copyright, trade and service marks and other proprietary notices are maintained. By accessing, sharing, receiving or otherwise using the Springer Nature journal content you agree to these terms of use (“Terms”). For these purposes, Springer Nature considers academic use (by researchers and students) to be non-commercial.

These Terms are supplementary and will apply in addition to any applicable website terms and conditions, a relevant site licence or a personal subscription. These Terms will prevail over any conflict or ambiguity with regards to the relevant terms, a site licence or a personal subscription (to the extent of the conflict or ambiguity only). For Creative Commons-licensed articles, the terms of the Creative Commons license used will apply.

We collect and use personal data to provide access to the Springer Nature journal content. We may also use these personal data internally within ResearchGate and Springer Nature and as agreed share it, in an anonymised way, for purposes of tracking, analysis and reporting. We will not otherwise disclose your personal data outside the ResearchGate or the Springer Nature group of companies unless we have your permission as detailed in the Privacy Policy.

While Users may use the Springer Nature journal content for small scale, personal non-commercial use, it is important to note that Users may not:

1. use such content for the purpose of providing other users with access on a regular or large scale basis or as a means to circumvent access control;
2. use such content where to do so would be considered a criminal or statutory offence in any jurisdiction, or gives rise to civil liability, or is otherwise unlawful;
3. falsely or misleadingly imply or suggest endorsement, approval, sponsorship, or association unless explicitly agreed to by Springer Nature in writing;
4. use bots or other automated methods to access the content or redirect messages
5. override any security feature or exclusionary protocol; or
6. share the content in order to create substitute for Springer Nature products or services or a systematic database of Springer Nature journal content.

In line with the restriction against commercial use, Springer Nature does not permit the creation of a product or service that creates revenue, royalties, rent or income from our content or its inclusion as part of a paid for service or for other commercial gain. Springer Nature journal content cannot be used for inter-library loans and librarians may not upload Springer Nature journal content on a large scale into their, or any other, institutional repository.

These terms of use are reviewed regularly and may be amended at any time. Springer Nature is not obligated to publish any information or content on this website and may remove it or features or functionality at our sole discretion, at any time with or without notice. Springer Nature may revoke this licence to you at any time and remove access to any copies of the Springer Nature journal content which have been saved.

To the fullest extent permitted by law, Springer Nature makes no warranties, representations or guarantees to Users, either express or implied with respect to the Springer nature journal content and all parties disclaim and waive any implied warranties or warranties imposed by law, including merchantability or fitness for any particular purpose.

Please note that these rights do not automatically extend to content, data or other material published by Springer Nature that may be licensed from third parties.

If you would like to use or distribute our Springer Nature journal content to a wider audience or on a regular basis or in any other manner not expressly permitted by these Terms, please contact Springer Nature at

[onlineservice@springernature.com](mailto:onlineservice@springernature.com)